

Comprehensive Analyses of Transport Proteins Encoded Within the Genome of “*Aromatoleum aromaticum*” Strain EbN1

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Abstract The denitrifying bacterium “*Aromatoleum aromaticum*” strain EbN1 is specialized for the aerobic utilization of aromatic compounds including crude oil constituents. We here report whole-genome analyses for potential transport proteins in *A. aromaticum* strain EbN1. This organism encodes very few transporters for simple sugars and most other common carbon sources. However, up to 28% of its putative transporters may act on fairly hydrophobic aromatic and aliphatic compounds. We categorize the putative transporters encoded within the genome, assign them to recognized families, and propose their preferred substrates. The bioinformatic data are correlated with available metabolic information to obtain an integrated view of the metabolic network of *A. aromaticum* strain EbN1. The results thus indicate that this organism possesses a disproportionately large percentage of transporters for the uptake and efflux of hydrophobic and amphipathic aromatic and aliphatic compounds compared with previously analyzed organisms. We predict that these

findings will have important implications for our eco-physiological understanding of bioremediation.

Keywords Anaerobic · Aromatic compounds · Degradation · Denitrification · Transport · Genome analysis

Introduction

Aromatic compounds are among the most widely encountered basic chemical structures in nature. They include protein constituents in all living organisms, abundant components in higher plants (e.g., the phenolic building blocks of lignin), and main constituents of crude oil (e.g., toluene). Fuel-derived alkylbenzenes and phenolic compounds are widely used in the chemical industry. Their accidental release into the environment often leads to accumulation in anoxic soils and sediments, increasingly demanding bioremediation efforts. The latter benefit from an in-depth understanding of relevant metabolic pathways and essential transport systems for their uptake and efflux at toxic concentrations.

Aerobic degraders of aromatic compounds and their metabolic pathways for initial hydroxylation and ring cleavage with O₂-dependent oxygenases have been known for decades (Harayama et al. 1992), but anaerobic degradation of these compounds has been discovered more recently. In fact, information concerning the involved enzyme mechanisms has been forthcoming only within the past decade (Gibson and Harwood 2002; Heider and Fuchs 1997). Since these enzymes cannot use species of reactive oxygen, their mechanisms differ fundamentally from those of the aerobic enzymes. Often they are found to involve unprecedented biochemistry (Gibson and Harwood 2002; Heider and Fuchs 1997) and provide valuable resources for

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biotechnological applications (Höffken et al. 2006; Wakett and Hershberger 2001).

The degradation of aromatic compounds usually involves channeling of the various phenolic, carboxylic, and hydrocarbon substrates into the pool of central intermediates, often benzoyl-CoA (Heider and Fuchs 1997). This intermediate subsequently undergoes reductive dearomatization and hydrolytic ring cleavage (Boll et al. 2002; Harwood and Gibson 1988). The microorganisms that degrade these compounds aerobically also use unusual pathways and novel types of oxygenases for aerobic aromatic compound metabolism (Mohamed Mel et al. 2002; Zaar et al. 2001).

Most currently known anaerobic aromatic compound-degrading bacteria are denitrifiers affiliated with the β -proteobacterial *Azoarcus/Thauera* cluster (Spormann and Widdel 2000; Widdel and Rabus 2001). This group is distinct from plant-associated *Azoarcus* spp. (Reinhold-Hurek and Hurek 2000) and has been proposed as the new genus *Aromatoleum* (Trautwein et al. 2008; Wöhlbrand et al. 2007, 2008; Wöhlbrand and Rabus 2008; Rabus, unpublished data). Distinction of the two genera is also reflected by the low degree of genome synteny (low frequency of colocalization of genes) between strain EbN1 and strain BH72 (Krause et al. 2006). *Aromatoleum aromaticum* strain EbN1 is a well-studied and metabolically versatile representative of this group (Rabus and Widdel 1995, 1996; Rabus, unpublished data). Strain EbN1 can grow anaerobically with alkylbenzenes, constituents of crude oil, polar aromatic compounds (e.g., phenol), and a variety of small polar aliphatic compounds (e.g., acetone).

The complete genome of strain EbN1 was the first to be determined for a member of the *Aromatoleum/Azoarcus/Thauera* cluster (Rabus et al. 2005). The genome consists of a single circular chromosome (4.3 Mb) and two plasmids (0.21 and 0.22 Mb) encoding a total of 4603 predicted proteins. The genome sequence revealed the genetic blueprint for a complex degradation network including more than 10 anaerobic pathways for aromatic compounds. Their absence in the aerobic aromatic compound-degrader *Pseudomonas putida* (Nelson et al. 2002) and the aerobic alkane-degrader *Alcanivorax borkumensis* (Schneiker et al. 2006) emphasizes the metabolic distinctiveness of strain EbN1. The only other sequenced genome of a bacterium similarly capable of anaerobic aromatic compound degradation is the phototrophic *Rhodopseudomonas palustris* (Larimer et al. 2004). The presence of multiple respiratory complexes and a large number of regulatory proteins suggests that strain EbN1 can adapt well to changing nutritional conditions.

In this report, we examine the complete complement of transporters encoded within the genome of *A. aromaticum*

strain EbN1 to decipher its key coordination points for solute exchange with the environment. We show that this organism encodes a larger percentage of putative transporters for the energy-driven uptake and export of aromatic/hydrophobic substances than any other previously examined organism. Potential substrates of some of these systems are identified, and the mechanisms of transport can often be deduced. The transport capabilities of this organism correlate with its previously determined degradative capacities.

Results

Overview of *Aromatoleum aromaticum* Strain EbN1 Transporters

Table 1 and Fig. 1 present an overview of transporters encoded within the *A. aromaticum* strain EbN1 genome. Two hundred fifty-eight transport systems were identified (see Table 3), many of them displaying multicomponent structures. These consist of 498 proteins, comprising 11% of the 4603 proteins predicted to be encoded in the genome of strain EbN1. This is comparable to the average fraction of predicted transporter proteins reported for other bacteria, although there is substantial variation (3%–17%) from organism to organism (Barabote and Saier 2005; Lorca et al. 2007; Ren and Paulsen 2007).

Forty-three of the 258 transport systems identified (17%) are simple channel proteins, either of the α (cytoplasmic membrane) type (11) or of the β (outer lipopolysaccharide-containing membrane) type (32). Secondary carriers are the most numerous type of transporter found in strain EbN1 (84 systems, or 33%), with primary active transporters coming in second place (68 systems, or 26%). This is not surprising for an organism with potential for both aerobic and anaerobic lifestyles. This organism has only one group translocating sugar transporter of the phosphotransferase system (PTS) already an indication that sugar metabolism is of little importance to strain EbN1 (see Fig. 1).

Strain EbN1 has a full complement of electron carrier complexes that couple transmembrane H^+ or Na^+ transport to electron flow (11 systems). There are also many transmembrane carriers that transfer electrons across the cytoplasmic membrane, from one side of the membrane to the other, thus influencing the energetics of the cell (16 systems). Sixteen auxiliary transport proteins (TC class 8) were identified which do not actually catalyze transport but facilitate transport processes catalyzed by recognized transporters. Additionally, there are 46 homologues of poorly defined systems, 8 of TC class 9A (transporters of

Table 1 Overview of the “*Aromatoleum aromaticum*” strain EbN1 transporter analyses

TC class ^a	Class description	No. of transport proteins	TC subclass	Subclass description	No. of transport proteins
1	Channels	43	1.A	α -type channels	11
			1.B	β -barrel porins	32
2	Secondary carriers	84	2.A	Porters (uniporters, symporters, antiporters)	81
			2.C	Ion-gradient-driven energizers	3
3	Primary active transporters	68	3.A	P-P-bond hydrolysis-driven transporters	57
			3.D	Oxidoreduction-driven transporters	11
4	Group translocators	1	4.A	Phosphotransfer-driven group translocators	1
5	Transmembrane electron carriers	16	5.A	Transmembrane 2-electron transfer carriers	12
				Transmembrane 1-electron transfer carrier	4
8	Auxillary transport protein ^b	[16]	8.A	Auxiliary transport proteins	[16]
9	Poorly defined system	46	9.A	Recognized transporters of unknown biochemical mechanism	8
			9.B	Putative uncharacterized transport proteins	38
Total		274			274

^a Transporter classes 6 and 7 have not been assigned in the TC system yet and therefore are not listed here

^b Auxillary proteins facilitate transport via established transport systems and therefore are not counted as separate systems

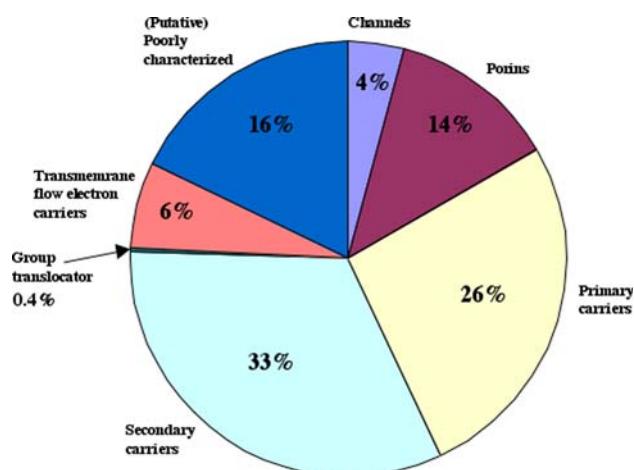


Fig. 1 Percentages of recognized transport systems found in *A. aromaticum* (formerly *Azoarcus*) strain EbN1 according to transporter type. “Channels” are of type 1A and β -barrel porins are of type 1B in the transporter classification database (TCDB). Percentages of secondary carriers (TC class 2), primary active transporters (class 3), group translocators (class 4), transmembrane electron carriers (class 5), and electron carriers that couple electron flow to H^+ or Na^+ extrusion (class 3D) are also presented. Poorly characterized transport proteins belong to TC class 9

unknown mechanisms of action) and 38 of TC class 9B (putative transporters). The latter group consists of a variety of potential transport systems, many families of which have not been characterized functionally. Finally, we have identified several distant homologues of recognized transporters with the expected topological features but unpredictable specificities (see below).

Potential α -Type Channel-Forming Proteins

A. aromaticum strain EbN1 encodes a single putative member of the chloride channel (CIC) family as well as a protein showing limited sequence similarity to members of the one-TMS organellar chloride channel (O-CIC) family. The latter protein resembles in sequence full-length glutathione S-transferases (GSTs), eukaryotic elongation factor 1- γ , HSP26 stress-related proteins, and a stringent starvation protein in *Escherichia coli*. The last three protein types mentioned are not known to have GST activity (Williams et al. 1994). It seems unlikely that the protein in strain EbN1 showing sequence similarity to O-CIC is a chloride channel.

Strain EbN1 has four homologues similar to mammalian gp91phox, a phagocyte NADPH oxidase with a cytochrome b558 domain (Kimball and Saier 2002). The latter suggestion has recently been questioned (Ramsey et al. 2006; Sasaki et al. 2006). This enzyme and its homologues (such as ferric oxidoreductases) have only been characterized in eukaryotes. However, the presence of full-length bacterial homologues has been recognized (Kimball and Saier 2002). Their functions are unknown, but functional parallels with the eukaryotic homologues suggest that these bacterial proteins catalyze transmembrane electron flow, from the cytoplasm into the periplasm. Three of the strain EbN1 homologues appear to be full length, but the fourth is much shorter, perhaps bearing only one or two of the domains of these multidomain proteins.

Strain EbN1 has one homologue of the MscL large-conductance mechanosensitive channel (MscL) family as

well as five members of the small-conductance mechanosensitive channel (MscS) family. These channel proteins in *E. coli* have been functionally characterized in terms of their conductance properties and their high-resolution X-ray structures (Chang et al. 1998; Martinac et al. 1987, 1990; Perozo et al. 2001; Sukharev et al. 1999, 2001). One of the MscS channels in strain EbN1 has 11 putative TMSs, while the rest have 6. In *E. coli*, these proteins are known to function in osmotic stress adaptation (Pivetti et al. 2003).

A MotA bacterial flagellar motor protein homologue (Hosking et al. 2006; Zhai et al. 2003), with four putative TMSs as expected, was found encoded in the genome of strain EbN1. Mapping at a distant location, a MotB homologue with the single expected TMS, which functions together with MotA, was found. These two proteins presumably function in strain EbN1 as the flagellar motor (Berry and Armitage 1999; DeRosier 1998).

Strain EbN1 has three DnaK protein homologues. The primary function of these protein chaperone homologues is in protein folding (Winter and Jakob 2004). In eukaryotes, DnaK proteins can insert into membranes to form cation-selective channels (Arispe and De Maio 2000). There is no evidence that these chaperone proteins form channels in prokaryotes.

Finally, strain EbN1 has a single CorA homologue of the heavy metal ion transporter (MIT) family (Table 2). These homopentameric channel proteins with two TMSs per subunit allow uptake of divalent cations such as Mg²⁺, Co²⁺, Ni²⁺, Zn²⁺, and Cd²⁺ (Kehres et al. 1998; Maguire 2006). The three-dimensional structure of the *E. coli* CorA has been solved (Maguire 2006).

In summary, strain EbN1 appears to have very few α -type channels in its cytoplasmic membrane. We are confident only that it has a single chloride channel of the CIC family and a single member of the MIT family of CorA proteins for uptake of divalent cations, as well as one mechanosensitive channel of the MscL family and five homologues of the MscS family. All of the Msc channels may function in hypoosmotic stress adaptation, possibly under different types of stress conditions.

Outer Membrane Porins

Strain EbN1 has a large number of homologues of recognized outer membrane porins (both general and substrate specific porins; Tables 1 and 2) (Nikaido 2003). These include six members of the 16-TMS General Bacterial Porin (GBP) family and three members of the 8-TMS OmpA-OprF Porin (OOP) family (Table 2). They also include single members of three other porin families that may exhibit substrate specificity. Thus, the 14-TMS FadL homologue may be specific for fatty acids and other hydrophobic substances (TC 1.B.9) (van den Berg et al.

2004), and the OprB homologue may be specific for sugars such as glucose (TC 1.B.19) (Wylie and Worobec 1995). Finally, the OmpW porin homologue may be specific for alkanes, hydrophobic drugs, and dyes such as methyl viologen (TC 1.B.39) (Santiviago et al. 2002; van Beilen et al. 1992). This conclusion is supported by structural studies (Albrecht R et al. 2006).

In addition to these more “traditional” porins, strain EbN1 has eight outer membrane receptors of the OMR family. Seven of these may function in the energy-dependent active uptake of iron-siderophore complexes across the outer membrane, but the eighth probably functions in vitamin B₁₂ uptake (Table 2). One of the seven putative siderophore receptors is truncated due to insertion of a Tnp30 (locus tag 63243; IS5 family) transposon, present in the genome of *A. aromaticum* strain EbN1.

There are five outer membrane factors (OMFs; TC 1.B.17) that normally function in a heterotrimeric complex for the efflux of proteins, drugs, and aromatic compounds via a pathway that involves a primary cytoplasmic membrane efflux pump and a largely periplasmic “membrane fusion protein” (MFP; TC 8.A.1). These systems allow efflux from the cytoplasm, through the cytoplasmic and outer membranes, to the external milieu in a single energy-coupled step (Dinh et al. 1994).

Strain EbN1 also has two outer membrane auxiliary (OMA) family members. These proteins appear to function exclusively in complex exo- or capsular polysaccharide export together with a primary cytoplasmic membrane transporter and other auxiliary proteins. They form octameric ring-like structures in the outer membrane with an inner diameter of about 2 nm (Beis et al. 2004; Nesper et al. 2003). One such protein, Wza, may be largely α -helical, an unusual but not totally novel property of an outer membrane porin (Beis et al. 2004; Collins et al. 2007; Dong et al. 2006; Nesper et al. 2003).

Like OMA family members, secretins form oligomeric (dodecameric) ring-like structures in the outer membrane (Collins et al. 2001, 2003). They function in conjunction with types II and III protein secretion systems (Collins et al. 2005). Strain EbN1 has three secretins. One is most similar to ComE of *Haemophilus influenzae*, a protein that functions in DNA uptake; the second is most similar to XcpQ of *Pseudomonas aeruginosa*; and the third most closely resembles Puld of *Klebsiella oxytoca*. Both of the latter two homologues are components of type II protein secretion (main terminal branch) systems (Saier 2006; Sandkvist 2001). As shown below, these three secretins may function with three dissimilar macromolecular transport systems.

Strain EbN1 has all four recognized constituents of the outer membrane protein insertion porin (OmpIP) family. It has one homologue of the *E. coli* YaeT protein, most

Table 2 Recognized transporters encoded within the genome of *Azoarcus* sp. EbN1

TC subject protein	Family name	No. of complete systems	EbN1ID no.	TC homologue (protein name)	EbN1 query length	TC subject length	E-value	No. of TMSS	Estimated transporter status
1.A.11.1.1	Ammonia Transporter Channel (Amt)	1	ebA3151	AmtB	428	451	E-126	12	Complete
1.A.11.2.3	Ammonia Transporter Channel (Amt)	1	ebA4609	AmtL	391	401	6.00E-38	11 (TC 12)	Complete
1.A.12.1.1	Organellar Chloride Channel (O-CIC)	1	ebA1195	CLIC5 orp64	198	437	2.00E-06	1	Complete
1.A.22.1.1	Large Conductance Mechanosensitive Ion Channel (MscL)	1	ebA2712	MscL	139	136	4.00E-26	2	Complete
1.A.23.1.1	Small Conductance Mechanosensitive Ion Channel (MscS)	5	ebA645	KefA	1120	8.00E-23	11	Complete	
1.A.23.4.1	Small Conductance Mechanosensitive Ion Channel (MscS)		ebA6926	MscMJ	1106	2.00E-05	5 (TC 6)	Complete	
1.A.23.4.2	Small Conductance Mechanosensitive Ion Channel (MscS)		ebA4035	MscMLR	350	1.00E-35	7 (TC 6)	Complete	
1.A.23.4.2	Small Conductance Mechanosensitive Ion Channel (MscS)		ebA4066	MscMLR	418	350	8.00E-08	6	Complete
1.A.23.4.2	Small Conductance Mechanosensitive Ion Channel (MscS)		ebA6709	MscMLR	278	350	3.00E-15	3 (TC 6)	Complete
1.A.30.1.4	H ⁺ - or Na ⁺ -translocating Bacterial Flagellar Motor (Mot)	1	ebA697	MotP	246	272	1.00E-39	4	Complete
1.A.30.1.4	H ⁺ - or Na ⁺ -translocating Bacterial Flagellar Motor (Mot)		ebA4407	MotS	190	261	2.00E-16	1	
1.A.35.3.1	CorA Metal Ion Transporter (MIT)	1	ebA4496	CorA	337	317	2.00E-20	3	Complete
1.A. total		11							
1.B.1.5.1	General Bacterial Porin (GBP)	1	p2A117	Omal	397	326	2.00E-17	1	Complete
1.B.1.6.1	General Bacterial Porin (GBP)	5	ebA1962	Porin protein 32	346	351	2.00E-36	1	Complete
1.B.1.6.1	General Bacterial Porin (GBP)		ebA2143	Porin protein 32	349	351	2.00E-36	1	Complete
1.B.1.6.1	General Bacterial Porin (GBP)		ebA2444	Porin protein 32	359	351	2.00E-31	1	Complete
1.B.1.6.1	General Bacterial Porin (GBP)		ebA3358	Porin protein 32	349	351	8.00E-32	1	Complete
1.B.1.6.1	General Bacterial Porin (GBP)		ebA5644	Porin protein 32	341	351	2.00E-34	0 (TC 1)	Complete
1.B.6.1.1	OmpA-OmpF Porin (OOP)	1	ebA913	OmpA	238	346	6.00E-32	1	Probably complete
1.B.6.1.2	OmpA-OmpF Porin (OOP)	1	ebA1121	OmpF	232	350	6.00E-24	3 (TC 1)	Probably complete
1.B.6.1.3	OmpA-OmpF Porin (OOP)	1	p1B164	OmpATb	202	326	5.00E-07	1	Probably complete
1.B.9.1.1	FadL Outer Membrane Protein (FadL)	1	c1A203	FadL	423	448	4.00E-50	1	Complete
1.B.14.1.2	Outer Membrane Receptor (OMR)	1	eba3939	FecA	695	774	4.00E-84	1	Complete
1.B.14.1.4	Outer Membrane Receptor (OMR)	2	eba5333	FhuA	700	747	3.00E-51	1	Complete
1.B.14.1.4	Outer Membrane Receptor (OMR)		eba3937	FhuA	699	747	2.00E-35	0 (TC 1)	Complete
1.B.14.1.6	Outer Membrane Receptor (OMR)	1	eba3244	CirA	278	663	3.00E-14	1 (TC 0)	Complete
1.B.14.2.3	Outer Membrane Receptor (OMR)	2	eba2117	HemR	686	687	9.00E-39	1 (TC 0)	Complete
1.B.14.2.3	Outer Membrane Receptor (OMR)		eba6096	HemR	713	687	3.00E-09	1 (TC 0)	Complete
1.B.14.3.1	Outer Membrane Receptor (OMR)	1	eba4013	BtuB	646	614	1.00E-17	1 (TC 0)	Complete
1.B.14.9.1	Outer Membrane Receptor (OMR)	1	eba6149	RhtA	703	746	1.00E-21	0	Complete
1.B.17.1.1	Outer Membrane Factor (OMF)	2	eba1642	TolC	453	495	3.00E-49	1	Complete
1.B.17.1.1	Outer Membrane Factor (OMF)		eba1783	TolC	483	495	2.00E-20	0 (TC 1)	Complete

Table 2 continued

TC subject protein	Family name	No. of complete systems	EbN1ID no.	TC homologue (protein name)	EbN1 query length	TC subject length	E-value	No. of TMSS	Estimated transporter status
1.B.17.3.2	Outer Membrane Factor (OMF)	1	ebA3445	FusA	475	530	3.00E-46	0 (TC 1)	Complete
1.B.17.3.3	Outer Membrane Factor (OMF)	1	ebA1847	OpeM	501	512	2.00E-33	0 (TC 1)	Complete
1.B.17.3.4	Outer Membrane Factor (OMF)	1	ebA1679	SilC	477	461	4.00E-40	1	Complete
1.B.18.3.1	Outer Membrane Auxiliary (OMA) Protein	2	ebA4247	Wza	211	379	3.00E-18	1	Complete
1.B.18.3.1	Outer Membrane Auxiliary (OMA) Protein		ebA6799	Wza	234	379	3.00E-08	0 (TC 1)	Complete
1.B.19.1.1	Glucose-selective OprB Porin (OprB)		ebA1863	OprB porin	494	454	1.00E-04	1	Complete
1.B.22.1.1	Outer Bacterial Membrane Secretin (Secretin)		ebA3713	PulD	566	660	8.00E-19	1	Probably complete
1.B.22.1.2	Outer Bacterial Membrane Secretin (Secretin)		ebA1602	XcpQ	770	658	1.00E-31	1	Complete
1.B.22.4.1	Outer Bacterial Membrane Secretin (Secretin)		ebA1239	ComE	771	445	5.00E-22	1	Complete
1.B.33.1.2	Outer Membrane Protein Insertion Porin (OmpIP)		ebA1189	D15	585	797	4.00E-05	1	Complete
1.B.33.1.3	Outer Membrane Protein Insertion Porin (OmpIP)		ebA773	NlpB	379	344	0.002	1	Complete
1.B.33.1.3	Outer Membrane Protein Insertion Porin (OmpIP)		ebA4724	YfiO	266	245	6.00E-37	1	Complete
1.B.33.1.3	Outer Membrane Protein Insertion Porin (OmpIP)		ebA1258	yfgL	381	392	5.00E-33	1	Complete
1.B.33.1.3	Outer Membrane Protein Insertion Porin (OmpIP)		ebA5996	yaeT	767	810	e-147	0	Complete
1.B.33.1.3	Outer Membrane Protein Insertion Porin (OmpIP)		ebA1141	OstA	787	784	2.00E-64	0	Complete
1.B.39.1.1	Bacterial Porin, OmpW (OmpW)	1	ebA5811	OmpW	225	212	8.00E-37	0 (TC 1)	Complete
1.B total			c2A195	Bcr	426	396	1.00E-36	12	Complete
2.A.1.2.7	Drug:H ⁺ Antipporter-1 (12 Spanner) (DHAI)	3	ebA3158	Bcr	394	396	6.00E-31	12	Complete
2.A.1.2.7	Drug:H ⁺ Antipporter-1 (12 Spanner) (DHAI)		ebA3906	Bcr	402	396	3.00E-13	12	Complete
2.A.1.2.7	Drug:H ⁺ Antipporter-1 (12 Spanner) (DHAI)		ebA3858	NorA	384	388	1.00E-13	12	Complete
2.A.1.2.10	Drug:H ⁺ Antipporter-1 (12 Spanner) (DHAI)	1	ebA3448	FarB	514	507	e-126	14	Complete
2.A.1.3.20	Drug:H ⁺ Antipporter-1 (14 Spanner) (DHAI)	1	ebA1213	NasA	404	421	8.00E-61	12 (TC 9)	Complete
2.A.1.8.2	Nitrate/Nitrite Porter (NNP)	1	ebA6289	NarK_N	458	905	3.00E-87	12 (TC 24)	Complete
2.A.1.8.11	Nitrate/H ⁺ symporter (NNP)	1	ebA6288	NarK_C	563	905	4.00E-57	14 (TC 24)	

Table 2 continued

TC subject protein	Family name	No. of complete systems	EbN1ID no.	TC homologue (protein name)	EbN1 query length	TC subject length	E-value	No. of TMSS	Estimated transporter status
2.A.1.11.1	Oxalate:Formate Antipporter (OFA)	1	ebA6311	OxIT	419	418	0.001	12	Probably complete
2.A.1.15.1	Aromatic Acid:H ⁺ Symporter (AAHS)	2	ebA4989	PcaK	467	448	7.00E-53	12	Complete
2.A.1.15.1	Aromatic Acid:H ⁺ Symporter (AAHS)		ebA5710	PcaK	399	448	6.00E-07	12	Complete
2.A.1.15.5	Aromatic Acid:H ⁺ Symporter (AAHS)	1	ebA5311	BenK	452	466	2.00E-97	12	Complete
2.A.1.17.1	Cyanate Porter (CP)	1	ebA3751	CynX	447	384	2.00E-25	11	Complete
2.A.1.17.1	Cyanate Porter (CP)		ebA725	CynX	415	384	1.00E-10	12 (TC 11)	Complete
2.A.1.25.2	Peptide-Acetyl-Coenzyme A Transporter (PAT)	1	ebA2813	AmpG	431	491	9.00E-65	12 (TC 14)	Complete
2.A.1.27.1	Phenyl Propionate Permease (PPP)	1	ebA4754	HeiT	386	379	3.00E-26	12	Complete
2.A.1.30.1	Putative Abietane Diterpenoid Transporter (ADT)	1	ebA5677	DitE	523	547	2.00E-72	12	Complete
2.A.1.36.1	Acriflavin-sensitivity (YnfM)	1	ebA5055	YnfM	419	417	6.00E-56	12	Complete
2.A.1.42.1	Lysophospholipid Transporter (LpIT)	2	ebA3471	LpIT	450	397	6.00E-63	12	Complete
2.A.1.42.1	Lysophospholipid Transporter (LpIT)		ebA6485	LpIT	416	397	1.00E-54	11 (TC 12)	Complete
2.A.1.46.1	Unknown Major Facilitator-5 (UMF5)	1	ebA2917	Transporter	402	396	4.00E-10	12	Complete
2.A.1.51.1	Unknown Major Facilitator 7 (UMF7)	1	ebA1383	Putative MFS permease	398	398	8.00E-164	12	Probably complete
2.A.4.1.4	Cation Diffusion Facilitator (CDF)	1	ebA4853	ZitB	331	313	8.00E-58	6 (TC 5)	Complete
2.A.4.1.5	Cation Diffusion Facilitator (CDF)	1	ebA4214	YipP	317	300	2.00E-27	5 (TC 6)	Complete
2.A.6.1.4	Heavy Metal Efflux (HME)	1	ebA2165	CusF	201	110	1.00E-05	0	Complete
2.A.6.1.4	Heavy Metal Efflux (HME)		ebA2169	CusC	434	457		1 (TC 2)	
2.A.6.1.4	Heavy Metal Efflux (HME)		ebA2168	CusB	520	407	8.00E-38	0 (TC 1)	
2.A.6.1.4	Heavy Metal Efflux (HME)		ebA2176	CusA	1058	1047	0	12	
2.A.6.2.7	(Largely Gram-negative Bacterial) Hydrophobe/Amphiphile Efflux-1 (HAEI)	1	ebA3873	AcrD	1083	1037	2.00E-59	13 (TC 12)	Complete
2.A.6.2.15	(Largely Gram-negative Bacterial) Hydrophobe/Amphiphile Efflux-1 (HAEI)	1	ebA7101	MexD	1024	1043	e-129	11 (TC 12)	Complete
2.A.6.2.16	(Largely Gram-negative Bacterial) Hydrophobe/Amphiphile Efflux-1 (HAEI)	1	ebA1649	MexE	368	414	2.00E-08	2 (TC 1)	Complete
2.A.6.2.16	(Largely Gram-negative Bacterial) Hydrophobe/Amphiphile Efflux-1 (HAEI)	1	ebA3924	MexE	334	414	1.00E-08	1 (TC 0)	Complete
2.A.6.2.17	(Largely Gram-negative Bacterial) Hydrophobe/Amphiphile Efflux-1 (HAEI)	2	ebA1647	MexK	1080	1025	0	12	Complete
2.A.6.2.17	(Largely Gram-negative Bacterial) Hydrophobe/Amphiphile Efflux-1 (HAEI)		ebA3926	MexK	1047	1025	3.00E-61	11 (TC 12)	Complete

Table 2 continued

TC subject protein	Family name	No. of complete systems	EbN1ID no.	TC homologue (protein name)	EbN1 query length	TC subject length	E-value	No. of TMSs	Estimated transporter status
2.A.6.2.21	(Largely Gram-negative Bacterial) Hydrophobe/Amphiphile Efflux-1 (HAE1)	1	ebA1973	OprM	475	485	3.00E-72	0	Complete
2.A.6.2.21	(Largely Gram-negative Bacterial) Hydrophobe/Amphiphile Efflux-1 (HAE1)		ebA1972	MexY	1054	1046	0	12	
2.A.6.2.21	(Largely Gram-negative Bacterial) Hydrophobe/Amphiphile Efflux-1 (HAE1)		ebA1971	MexX	404	389	7.00E-48	0	
2.A.6.7.1	(Largely Archaeal Putative Hydrophobe/ Amphiphile Efflux-3 (HAE3))	1	ebA5169	ORF in Afu	787	750	2.00E-22	12	Complete
2.A.6.7.2	(Largely Archaeal Putative Hydrophobe/ Amphiphile Efflux-3 (HAE3))	6	ebA327	ORF in Mja	799	388	8.00E-13	12	Complete
2.A.6.7.2	(Largely Archaeal Putative Hydrophobe/ Amphiphile Efflux-3 (HAE3))		c1A90	ORF in Mja	808	388	5.00E-11	12 (TC 6)	Probably complete
2.A.6.7.2	(Largely Archaeal Putative Hydrophobe/ Amphiphile Efflux-3 (HAE3))		ebA1928	ORF in Mja	799	388	4.00E-13	12 (TC 6)	Probably complete
2.A.6.7.2	(Largely Archaeal Putative Hydrophobe/ Amphiphile Efflux-3 (HAE3))		ebA4709	ORF in Mja	791	388	6.00E-12	12 (TC 6)	Probably complete
2.A.6.7.2	(Largely Archaeal Putative Hydrophobe/ Amphiphile Efflux-3 (HAE3))		ebA5763	ORF in Mja	807	388	3.00E-14	12 (TC 6)	Probably complete
2.A.6.7.2	(Largely Archaeal Putative Hydrophobe/ Amphiphile Efflux-3 (HAE3))		p2A399	ORF in Mja	791	388	4.00E-17	12 (TC 6)	Probably complete
2.A.6.8.1	Brominated Aryl Polyene Pigment Exporter (ORt4)	1	ebA7066	Putative inner membrane protein	783	807	2.00E-76	12	Complete
2.A.7.1.3	4 TMS Small Multidrug Resistance (SMR)	1	c1A283	EmrE	122	110	3.00E-10	4	Complete
2.A.7.3.2	TMS Drug/Metabolite Exporter (DME)	3	ebA2542	YdeD	289	299	1.00E-07	9 (TC 10)	Complete
2.A.7.3.2	10 TMS Drug/Metabolite Exporter (DME)		ebA3672	YdeD	294	299	2.00E-06	10	Complete
2.A.7.3.2	10 TMS Drug/Metabolite Exporter (DME)		ebA4612	YdeD	306	299	6.00E-09	10	Complete
2.A.7.3.4	10 TMS Drug/Metabolite Exporter (DME)	1	ebA6529	YwiM	297	297	1.00E-06	10	Complete
2.A.7.3.7	10 TMS Drug/Metabolite Exporter (DME)	1	ebA4959	Sam (RPO76)	318	294	8.00E-18	10	Complete
2.A.9.3.1	Cytochrome Oxidase Biogenesis (Oxa1)	1	ebA2842	YidC	550	548	e-114	4 (TC 3)	Complete
2.A.16.2.1	Tellurite-resistance/Dicarboxylate Transporter (TDT)	1	ebA1993	MaeI	358	438	4.00E-06	10	Complete
2.A.20.2.4	Inorganic Phosphate Transporter (PiT)	1	ebA6176	Pht2;1	527	587	5.00E-45	11 (TC 13)	Complete
2.A.21.7.1	Solute:Sodium Symporter (SSS)	1	ebA3695	Phenylacetate permease Ppa	365	520	1.00E-60	9 (TC 13)	Complete
2.A.21.7.2	Solute:Sodium Symporter (SSS)	2	ebA165	ActP	693	549	1.00E-23	14	Complete
2.A.21.7.2	Solute:Sodium Symporter (SSS)		ebA5537	ActP	572	549	0	14	Complete
2.A.21.8.1	Solute:Sodium Symporter (SSS)	1	ebA3608	CHT1	488	580	2.00E-37	13	Complete
2.A.21.9.2	Solute:Sodium Symporter (SSS)	1	ebA169	PhS	918	1156	e-103	13	Complete

Table 2 continued

TC subject protein	Family name	No. of complete systems	EbN1ID no.	TC homologue (protein name)	EbN1 query length	TC subject length	E-value	No. of TMSS	Estimated transporter status
2.A.25.1.1	Alanine or Glycine:Cation Symporter (AGCS)	1	ebA3983	DagA	475	542	1.00E-58	11	Complete
2.A.36.4.3	Monovalent Cation:Proton Antiporter-1 (CPA1)	1	ebA6098	Nhal	445	468	0.001	13	Complete
2.A.37.1.1	Monovalent Cation:Proton Antiporter-2 (CPA2)	1	ebA1314	KetC	656	620	2.00E-50	12	Complete
2.A.37.1.1	Monovalent Cation:Proton Antiporter-2 (CPA2)	1	ebA1313	YabF (ancillary protein)	176			0	
2.A.38.1.1	K ⁺ Transporter (Trk)	1	eba2824	TrkH	485	483	e-103	10 (TC 11)	Complete
2.A.38.1.1	K ⁺ Transporter (Trk)	1	eba2825	TrkA	467	458	e-133	1 (TC 0)	
2.A.38.1.1	K ⁺ Transporter (Trk)	2	eba2742	KirA	231	222	6.00E-40	1 (TC 0)	Complete
2.A.38.4.3	K ⁺ Transporter (Trk)	1	eba2741	KirB	444	445	7.00E-69	12 (TC 9)	
2.A.38.4.3	K ⁺ Transporter (Trk)	1	eba5556	KirA	231	222	6.00E-40	1 (TC 0)	Complete
2.A.38.4.3	K ⁺ Transporter (Trk)	1	eba5555	KirB	444	455	4.00E-53	12	
2.A.49.5.1	Chloride Channel (CIC)	1	eba4866	ErIC	468	473	1.00E-18	10 (TC 11)	Complete
2.A.51.1.2	Chromate Ion Transporter (CHR)	1	eba4644	SrpC	183	393	6.00E-13	5 (TC 12)	Probably complete
2.A.51.1.2	Chromate Ion Transporter (CHR)	1	eba4646	SrpC	164	393	8.00E-05	5 (TC 12)	
2.A.53.3.1	Sulfate Permease (Sulp)	1	eba4402	Sulfate permease	562	492	1.00E-48	10 (TC 13)	Complete
2.A.53.4.1	Sulfate Permease (Sulp)	1	eba4484	Sulfate transporter	570	566	1.00E-46	14 (TC 13)	Complete
2.A.56.1.1	Tripartite ATP-independent Periplasmic Transporter (TRAP-T)	1	eba4162	DctM	427	440	e-171	13	Complete
2.A.56.1.1	Tripartite ATP-independent Periplasmic Transporter (TRAP-T)	1	eba4159	DctQ	238	227	2.00E-60	4	
2.A.56.1.1	Tripartite ATP-independent Periplasmic Transporter (TRAP-T)	1	eba4158	DctP	334	333	e-125	1	
2.A.56.1.1	Tripartite ATP-independent Periplasmic Transporter (TRAP-T)	1	eba5367	DctM	439	440	1.00E-28	12 (TC 13)	Complete
2.A.56.1.1	Tripartite ATP-independent Periplasmic Transporter (TRAP-T)	1	eba5362	DctP	349	333	2.00E-15	1	
2.A.56.1.1	Tripartite ATP-independent Periplasmic Transporter (TRAP-T)	1	eba7188	DctQ	227	173		4	
2.A.56.1.1	Tripartite ATP-independent Periplasmic Transporter (TRAP-T)	1	eba729	DctP	344	333	7.00E-10	1	Complete
2.A.56.1.1	Tripartite ATP-independent Periplasmic Transporter (TRAP-T)	1	eba732	DctM	443	440	7.00E-52	12 (TC 13)	
2.A.56.1.1	Tripartite ATP-independent Periplasmic Transporter (TRAP-T)	1	eba733	DctQ	214	227	2.00E-04	4	
2.A.56.1.2	Tripartite ATP-independent Periplasmic Transporter (TRAP-T)	1	eba4994	YiaO (R)	343	328	3.00E-10	1	Complete
2.A.56.1.2	Tripartite ATP-independent Periplasmic Transporter (TRAP-T)	1	p2A96	4TMS + YiaN (M)	656	425	7.00E-13	18 (TC 12+4)	

Table 2 continued

TC subject protein	Family name	No. of complete systems	EbN1ID no.	TC homologue (protein name)	EbN1 query length	TC subject length	E-value	No. of TMSS	Estimated transporter status
2.A.56.1.5	Tripartite ATP-independent Periplasmic Transporter (TRAP-T)	C(3)	ebA1032	RRC01191 (mannitol-binding)	361	365	1.00E-101	1	Incomplete
2.A.56.1.5	Tripartite ATP-independent Periplasmic Transporter (TRAP-T)	1	ebA1033	RRC01191 (mannitol-binding)	361	365	1.00E-105	1	Complete
2.A.56.1.5	Tripartite ATP-independent Periplasmic Transporter (TRAP-T)	ebA1036	RRC02478 (S)	203	190	1.00E-44	4		
2.A.56.1.5	Tripartite ATP-independent Periplasmic Transporter (TRAP-T)	ebA1037	RRC02479 (L)	564	611	e-144	16 (TC 18)		
2.A.59.1.2	Arsenical Resistance-3 (ACR3)	1	ebA5001	ArkB	371	346	2.00E-46	10	Complete
2.A.63.1.1	Monovalent Cation (K^+ or Na^+):Proton Antipporter-3 (CPA3)	1	ebA553	PhaG	103	121	1.00E-09	3	Complete
2.A.63.1.1	Monovalent Cation (K^+ or Na^+):Proton Antipporter-3 (CPA3)	ebA554	PhaF	93	92	5.00E-23	3		
2.A.63.1.1	Monovalent Cation (K^+ or Na^+):Proton Antipporter-3 (CPA3)	ebA555	PhaE	160	161	1.00E-27	3		
2.A.63.1.1	Monovalent Cation (K^+ or Na^+):Proton Antipporter-3 (CPA3)	ebB15	PhaC	115	115	6.00E-34	3		
2.A.63.1.1	Monovalent Cation (K^+ or Na^+):Proton Antipporter-3 (CPA3)	ebA558	PhaA + PhaB	967	725	0	19		
2.A.63.1.1	Monovalent Cation (K^+ or Na^+):Proton Antipporter-3 (CPA3)	ebA556	PhaD	538	547	1.00E-93	14 (TC 17)		
2.A.64.1.1	Twin Arginine Targeting (Tat)	1	ebB38	TatE/TatA	75	67	4.00E-08	1	Complete
2.A.64.1.1	Twin Arginine Targeting (Tat)	ebD2	TatB	147	171	1.00E-05	1		
2.A.64.1.1	Twin Arginine Targeting (Tat)	ebA1285	TatC	262	258	4.00E-56	6 (TC 5)		
2.A.66.1.1	Multifunctional Extrusion (MATE)	1	ebA6316	NorM	463	456	2.00E-69	12	Complete
2.A.66.2.7	Polysaccharide Transport (PST)	1	ebAA295	WzcC	486	492	1.00E-20	14	Complete
2.A.66.4.1	Mouse Virulence Factor (MVF)	1	ebA5091	MviN	530	524	e-142	14	Complete
2.A.69.2.1	Auxin Efflux Carrier (AEC)	2	ebA2636	MdcF	291	319	5.00E-08	10	Complete
2.A.69.2.1	Auxin Efflux Carrier (AEC)	ebA5059	MdcF	313	319	2.00E-14	10	Complete	
2.A.71.2.1	Folate-Biotin Transporter (FBT)	1	ebA826	Synechococcus	541	453	8.00E-11	14	Complete
2.A.72.1.1	K^+ Uptake Permease (KUP)	1	ebA3621	KUP	640	622	e-160	11 (TC 13)	Complete
2.A.72.1.1	K^+ Uptake Permease (KUP)	ebD71	KUP	82	622	4.00E-07	0 (TC 13)		
2.A.76.1.1	Resistance to Homoserine/Threonine (RhtB)	1	ebA2598	RhtB	211	206	2.00E-11	6	Complete
2.A.76.1.2	Resistance to Homoserine/Threonine (RhtB)	1	ebA6561	RhtC	211	206	2.00E-22	5 (TC 6)	Complete
2.A.86.1.1	Putative Permease (PerM)	1	ebA7105	PerM	378	353	2.00E-19	8 (TC 7)	Complete
2.A total		81							

Table 2 continued

TC subject protein	Family name	No. of complete systems	EbN1ID no.	TC homologue (protein name)	EbN1 query length	TC subject length	E-value	No. of TMSs	Estimated transporter status
2.C.1.1	TonB-ExbB-ExbD/TolA-TolQ-TolR (TonB) of Auxiliary Proteins for Energization of Outer Membrane Receptor (OMR)-mediated Active Transport	2	ebA1839	TonB	230	239	3.00E-06	0 (TC 1)	Complete
2.C.1.1.1	TonB-ExbB-ExbD/TolA-TolQ-TolR (TonB) of Auxiliary Proteins for Energization of Outer Membrane Receptor (OMR)-mediated Active Transport		ebA1841	ExbB	259	230	1.00E-26	3	
2.C.1.1.1	TonB-ExbB-ExbD/TolA-TolQ-TolR (TonB) of Auxiliary Proteins for Energization of Outer Membrane Receptor (OMR)-mediated Active Transport		ebA1842	ExbD	138	142	3E-13	1	
2.C.1.1.1	TonB-ExbB-ExbD/TolA-TolQ-TolR (TonB) of Auxiliary Proteins for Energization of Outer Membrane Receptor (OMR)-mediated Active Transport		ebA5081	ExbB	202	244	2E-10	3	Complete
2.C.1.1.1	TonB-ExbB-ExbD/TolA-TolQ-TolR (TonB) of Auxiliary Proteins for Energization of Outer Membrane Receptor (OMR)-mediated Active Transport		ebA5156	TonB	265	239	3.00E-04	0 (TC 1)	
2.C.1.1.1	TonB-ExbB-ExbD/TolA-TolQ-TolR (TonB) of Auxiliary Proteins for Energization of Outer Membrane Receptor (OMR)-mediated Active Transport		ebB176	ExbD	141	142	142	1	
2.C.1.1.1	TonB-ExbB-ExbD/TolA-TolQ-TolR (TonB) of Auxiliary Proteins for Energization of Outer Membrane Receptor (OMR)-mediated Active Transport		ebA2644	TolQ	224	230	2E-53	3	Complete
2.C.1.2.1	TonB-ExbB-ExbD/TolA-TolQ-TolR (TonB) of Auxiliary Proteins for Energization of Outer Membrane Receptor (OMR)-mediated Active Transport	1	ebA2650	TolB	426	430	2.00E-74	1	
2.C.1.2.1	TonB-ExbB-ExbD/TolA-TolQ-TolR (TonB) of Auxiliary Proteins for Energization of Outer Membrane Receptor (OMR)-mediated Active Transport		ebA2645	TolR	137	141	2.00E-11	1	

Table 2 continued

TC subject protein	Family name	No. of complete systems	EbN1ID no.	TC homologue (protein name)	EbN1 query length	TC subject length	E-value	No. of TMSs	Estimated transporter status
2.C.1.2.1	TonB-ExbB-ExbD/TolA-TolQ-TolR (TonB) of Auxiliary Proteins for Energization of Outer Membrane Receptor (OMR)-mediated Active Transport		ebA2648	TolA/TolB			5.00E-11	1	
2.C.1.2.1	TonB-ExbB-ExbD/TolA-TolQ-TolR (TonB) of Auxiliary Proteins for Energization of Outer Membrane Receptor (OMR)-mediated Active Transport		ebA2652	Pal	175	173	2.00E-19	0 (TC 1)	
2.C.1.2.1	TonB-ExbB-ExbD/TolA-TolQ-TolR (TonB) of Auxiliary Proteins for Energization of Outer Membrane Receptor (OMR)-mediated Active Transport		ebA2654	YbgF	246	263	3.00E-18	0 (TC 1)	
2.C total		3	ebA4561	GltU (R)	298	302	8.00E-78	1	Complete
3.A.1.3.4	Polar Amino Acid Uptake Transporter (PAAT)	1	ebA4562	GltK (M)	226	224	3.00E-57	5	
3.A.1.3.4	Polar Amino Acid Uptake Transporter (PAAT)		ebA4564	GltJ (M)	247	246	1.00E-50	5	
3.A.1.3.4	Polar Amino Acid Uptake Transporter (PAAT)		ebA4567	GltL (C)	242	241	5.00E-99	0	
3.A.1.4.1	Hydrophobic Amino Acid Uptake Transporter (HAAT)	1	ebA3394	LivG (C)	241	255	1.00E-27	0	Complete
3.A.1.4.1	Hydrophobic Amino Acid Uptake Transporter (HAAT)		ebA2070	LivF (C)					
3.A.1.4.1	Hydrophobic Amino Acid Uptake Transporter (HAAT)		ebA2700	LivJ (R)	382	454	3.00E-15	1	
3.A.1.4.1	Hydrophobic Amino Acid Uptake Transporter (HAAT)		ebA2706	LivM (M)	321	425	1.00E-15	11 (TC 10)	
3.A.1.4.1	Hydrophobic Amino Acid Uptake Transporter (HAAT)		ebA3167	LivF	250	237	6.00E-55	0	Complete
3.A.1.4.1	Hydrophobic Amino Acid Uptake Transporter (HAAT)		ebA3169	LivG	258	255	5.00E-55	0	
3.A.1.4.1	Hydrophobic Amino Acid Uptake Transporter (HAAT)		ebA3170	LivM	366	425	4.00E-58	8 (TC 10)	
3.A.1.4.1	Hydrophobic Amino Acid Uptake Transporter (HAAT)		ebA3172	LivH	310	308	2.00E-43	9	

Table 2 continued

TC subject protein	Family name	No. of complete systems	EbN1 ID no.	TC homologue (protein name)	EbN1 query length	TC subject length	E-value	No. of TMSS	Estimated transporter status
3.A.1.4.1	Hydrophobic Amino Acid Uptake Transporter (HAAT)		ebA3173	LivK	372	369	5.00E-41	0 (TC 1)	
3.A.1.4.1	Hydrophobic Amino Acid Uptake Transporter (HAAT)		ebA3175	BraC	370	381	5.00E-50	1	
3.A.1.4.1	Hydrophobic Amino Acid Uptake Transporter (HAAT)	1	ebA3556	LivK	382	454	7.00E-16	1	Complete
3.A.1.4.1	Hydrophobic Amino Acid Uptake Transporter (HAAT)		ebA3558	LivH	345	308	1.00E-12	9	
3.A.1.4.1	Hydrophobic Amino Acid Uptake Transporter (HAAT)		ebA3559	LivG + LivM	600	255	1.00E-43	7 (TC 0)	
3.A.1.4.1	Hydrophobic Amino Acid Uptake Transporter (HAAT)		ebA3560	LivF	259	233	9E-54	0	
3.A.1.4.1	Hydrophobic Amino Acid Uptake Transporter (HAAT)		ebA3316	LivJ	380	367	8.00E-09	0 (TC 1)	
3.A.1.4.2	Hydrophobic Amino Acid Uptake Transporter (HAAT)	1	ebA3425	NatC	309	331	2.00E-14	8 (TC 9)	Complete
3.A.1.4.2	Hydrophobic Amino Acid Uptake Transporter (HAAT)		ebA3426	NatD	304	290	5.00E-19	7	
3.A.1.4.2	Hydrophobic Amino Acid Uptake Transporter (HAAT)		ebA3428	NatB	405	434	8.00E-08	0 (TC 1)	
3.A.1.4.2	Hydrophobic Amino Acid Uptake Transporter (HAAT)		ebA3429	NatE	243	237	2.00E-40	0	
3.A.1.4.2	Hydrophobic Amino Acid Uptake Transporter (HAAT)		ebA3430	NatA	251	267	2.00E-44	0 (TC 1)	
3.A.1.4.4	Hydrophobic Amino Acid Uptake Transporter (HAAT)	1	ebA5303	UrtA	395	434	2.00E-11	2 (TC 1)	Complete
3.A.1.4.4	Hydrophobic Amino Acid Uptake Transporter (HAAT)		ebA5304	UrtB	288	294	7.00E-26	7 (TC 8)	
3.A.1.4.4	Hydrophobic Amino Acid Uptake Transporter (HAAT)		ebA5306	UrtC	324	359	5E-23	8 (TC 9)	
3.A.1.4.4	Hydrophobic Amino Acid Uptake Transporter (HAAT)		ebA5307	UrtD	253	242	9E-42	0	
3.A.1.4.4	Hydrophobic Amino Acid Uptake Transporter (HAAT)		ebA5309	UrtE	238	237	7.00E-41	0	
3.A.1.5.2	Peptide/Opine/Nickel Uptake Transporter (PepT)	1	ebA6466	DppD (C)	660	335	4.00E-74	0	Probably complete
3.A.1.5.2	Peptide/Opine/Nickel Uptake Transporter (PepT)		ebA6658	DppC (M)	476	320	2.00E-29	9 (TC 6)	
3.A.1.5.2	Peptide/Opine/Nickel Uptake Transporter (PepT)		ebA6661	DppB (M)	325	308	3.00E-29	6	
3.A.1.5.2	Peptide/Opine/Nickel Uptake Transporter (PepT)		ebA6662	MppA (R)	723	543	7.00E-11	1	
3.A.1.5.2	Peptide/Opine/Nickel Uptake Transporter (PepT)		DppA (C)		274				

Table 2 continued

TC subject protein	Family name	No. of complete systems	EbN1 ID no.	TC homologue (protein name)	EbN1 query length	TC subject length	E-value	No. of TMSS	Estimated transporter status
3.A.1.5.11	Peptide/Opine/Nickel Uptake Transporter (PepT)	1	ebA5654	YliA	703	629	5E-93	0	Complete
3.A.1.5.11	Peptide/Opine/Nickel Uptake Transporter (PepT)		ebA5655	YliC (M)	317	306	1.00E-40	6	
3.A.1.5.11	Peptide/Opine/Nickel Uptake Transporter (PepT)		ebA5658	YliD	304	303	9.00E-35	7 (TC 5)	
3.A.1.5.11	Peptide/Opine/Nickel Uptake Transporter (PepT)		ebA5660	YliB (precursor)	527	512	1.00E-15	1 (TC 0)	
3.A.1.6.1	Sulfate/Tungstate Uptake Transporter (SutT)	1	ebA6204	SUBI (R)	333	329	e-102	0	Probably complete
3.A.1.6.1	Sulfate/Tungstate Uptake Transporter (SutT)		ebA6206	CysT (M)	299	277	4.00E-64	6	
3.A.1.6.1	Sulfate/Tungstate Uptake Transporter (SutT)		ebA6207	CysW (M)	299	291	8.00E-75	6	
3.A.1.6.1	Sulfate/Tungstate Uptake Transporter (SutT)		ebA6208	CysA (C)	354	365	9.00E-87	0 (TC 1)	
3.A.1.6.1	Could be ebB221 (67 aa) but does not hit a TC sequence?			CysP (R)		338			
3.A.1.6.2	Sulfate/Tungstate Uptake Transporter (SutT)	1	ebA3597	TupA	273	286	2.00E-62	1	Complete
3.A.1.6.2	Sulfate/Tungstate Uptake Transporter (SutT)		ebA5014	TupC	237	214	7.00E-34	0	
3.A.1.6.2	Sulfate/Tungstate Uptake Transporter (SutT)		ebA5015	TupB	246	228	3.00E-30	6 (TC 5)	
3.A.1.7.1	Phosphate Uptake Transporter (PhoT)	1	ebA4826	PstS	330	346	3.00E-06	0 (TC 1)	Complete
3.A.1.7.1	Phosphate Uptake Transporter (PhoT)		ebA4828	PstC	319	7.00E-24		6	
3.A.1.7.1	Phosphate Uptake Transporter (PhoT)		ebA4829	PstA	307	296	8.00E-23	6	
3.A.1.7.1	Phosphate Uptake Transporter (PhoT)		ebA4830	PstB	289	257	3.00E-81	0	
3.A.1.8.1	Molybdate Uptake Transporter (MolT)	1	ebA2719	ModC (C)	377	352	2.00E-65	0 (TC 1)	Complete
3.A.1.8.1	Molybdate Uptake Transporter (MolT)		ebA2720	ModB (M)	224	229	5.00E-18	5	
3.A.1.8.1	Molybdate Uptake Transporter (MolT)		ebA2721	ModA (R)	255	257	2.00E-13	1	
3.A.1.10.1	Ferric Iron Uptake Transporter (FeT)	1	ebA6521	StuA	374	338		1	Probably complete
3.A.1.10.1	Ferric Iron Uptake Transporter (FeT)		ebA6523	StuC		345		0 (TC 1)	
3.A.1.10.1	Ferric Iron Uptake Transporter (FeT)		ebA6524	StuB		527		7 (TC 12)	
3.A.1.10.2	Ferric Iron Uptake Transporter (FeT)	1	ebA4893	FutB	560	557	e-123	12	Complete
3.A.1.10.2	Ferric Iron Uptake Transporter (FeT)		ebA4918	FutA1/FutA2	339	346	1.00E-82	0 (TC 1)	
3.A.1.10.2	Ferric Iron Uptake Transporter (FeT)		ebA4892	FutC	372	368	2.00E-63	1 (TC 0)	
3.A.1.10.2	Ferric Iron Uptake Transporter (FeT)	C(4)	ebA2111	FutC	360	368	2.00E-60	Incomplete	
3.A.1.13.1	Vitamin B12 Uptake Transporter (B12T)	1 (similar to 3.A.1.14)	ebA3688	BtuF	297	266	1.00E-26	0 (TC 1)	Complete
3.A.1.13.1	Vitamin B12 Uptake Transporter (B12T)	(similar to 3.A.1.14)	ebA4019	BtuC	326	326	4.00E-15	10 (TC 9)	

Table 2 continued

TC subject protein	Family name	No. of complete systems	EbN1ID no.	TC homologue (protein name)	EbN1 query length	TC subject length	E-value	No. of TMSS	Estimated transporter status
3.A.1.13.1	Vitamin B12 Uptake Transporter (B12T) (similar to 3.A.1.14)		ebA4020	BtuD	259	249	8.00E-34	0	
3.A.1.14.3	Iron Chelate Uptake Transporter (FeCT)	C(3)	ebA3857	FhuC	957	265	7.00E-10	0	CDD-PRK0349, uvrA, exinuclease ABC subunit A
3.A.1.15.2	Manganese/Zinc/Iron Chelate Uptake Transporter (MZT) (similar to 3.A.1.12 & 3.A.1.16)	1	ebA1813	Seab	289	278	1.00E-26	7	Complete
3.A.1.15.2	Manganese/Zinc/Iron Chelate Uptake Transporter (MZT) (similar to 3.A.1.12 & 3.A.1.16)		ebA1814	Seaa	303	305	2.00E-48	1	
3.A.1.15.2	Manganese/Zinc/Iron Chelate Uptake Transporter (MZT) (similar to 3.A.1.12 & 3.A.1.16)		ebA1812	Seac	255	266	6.00E-37	0	
3.A.1.17.1	Taurine Uptake Transporter (TauT) (similar to 3.A.1.12 & 3.A.1.16)	2	ebA2113	TauB (C)	285	255	2.00E-34	0 (TC 1)	Complete
3.A.1.17.1	Taurine Uptake Transporter (TauT) (similar to 3.A.1.12 & 3.A.1.16)		ebA2114	TauC (M)	265	275	1.00E-10	6	
3.A.1.17.1	Taurine Uptake Transporter (TauT) (similar to 3.A.1.12 & 3.A.1.16)		ebA2115	TauA (R)	338	320	0.002	0	
3.A.1.17.1	Taurine Uptake Transporter (TauT) (similar to 3.A.1.12 & 3.A.1.16)		p2A122	TauC (M)	269	275	2.00E-35	6	Complete
3.A.1.17.1	Taurine Uptake Transporter (TauT) (similar to 3.A.1.12 & 3.A.1.16)		p2A128	TauA (R)	348	320	5.00E-07	1 (TC 0)	
3.A.1.17.1	Taurine Uptake Transporter (TauT) (similar to 3.A.1.12 & 3.A.1.16)		p2A125	TauB (C)	285	255	7.00E-32	0 (TC 1)	
3.A.1.102.1	Lipoooligosaccharide Exporter (LOSE)	4	ebA3579	NodI (M)	263	262	3.00E-54	6	Complete
3.A.1.102.1	Lipoooligosaccharide Exporter (LOSE)		ebA3580	NodI (C)	325	347	8.00E-85	0	
3.A.1.102.1	Lipoooligosaccharide Exporter (LOSE)		ebA6231	NodI (M)	253	262	2.00E-13	6	Complete
3.A.1.102.1	Lipoooligosaccharide Exporter (LOSE)		ebA6230	NodI (C)	311	325	1.00E-39	1 (TC 0)	
3.A.1.102.1	Lipoooligosaccharide Exporter (LOSE)		ebA6744	NodII + NodJ (CM)	972	347	3.00E-39	6 (TC 0)	Complete
3.A.1.103.1	Lipopolyssaccharide Exporter (LPSE)		ebA1592	RfbA (M)	273	259	6.00E-13	6	Complete
3.A.1.103.1	Lipopolyssaccharide Exporter (LPSE)		ebA1593	RfbB (C)	483	246	9.00E-53	0	
3.A.1.105.2	Drug Exporter-1 (DrugE1)	2	ebA6267	OleC4 (C)	317	325	1.00E-36	0	Complete
3.A.1.105.2	Drug Exporter-1 (DrugE1)		ebA6745	OleC5 (M)	393	273	2.00E-08	7 (TC 6)	
3.A.1.105.2	Drug Exporter-1 (DrugE1)		ebA1306	OleC4 (C)	298	325	4.00E-39	0	Complete
3.A.1.105.2	Drug Exporter-1 (DrugE1)		ebA1304	OleC5 (M)	256	262	1.00E-07	6	
3.A.1.106.1	Lipid Exporter (LipdE)	1	ebA3992	MsbA	601	582	e-122	6 (TC 5)	Complete
3.A.1.107.1	Putative Heme Exporter (HemeE)	1	ebA3514	CycV (C)	214	200	8.00E-26	0	Complete
3.A.1.107.1	Putative Heme Exporter (HemeE)		ebA3516	CycW (M)	225	222	1.00E-04	6	

Table 2 continued

TC subject protein	Family name	No. of complete systems	EbN1 ID no.	TC homologue (protein name)	EbN1 query length	TC subject length	E-value	No. of TMSS	Estimated transporter status
3.A.1.107.1	Putative Heme Exporter (HemeE)		ebA3518	CyZ (M)	257	263	1.00E-43	6	Complete
3.A.1.109.1	Protein-1 Exporter (ProtIE)	1	ebA1666	HlyB	1021	707	c-124	6 (TC 8)	Complete
3.A.1.109.3	Protein-1 Exporter (ProtIE)	1	ebA1807	LapB	742	718	e-156	6	Complete
3.A.1.110.9	Protein-2 Exporter (Prot2E)	1	ebA7181	HasD (M-C)	574	580	e-108	6	Complete
3.A.1.114.1	Probable Glycolipid Exporter (DevE)	1	ebA6251	DevC (M)	401	385	2.00E-05	4 (TC 6)	Complete
3.A.1.114.1	Probable Glycolipid Exporter (DevE)		ebA6253	DevA (C)	237	244	1.00E-36	0	Complete
3.A.1.120.1	(Putative) Drug Resistance ATPase-1 (Drug RA1)	1	ebA2533	SrmB	655	550	1.00E-51	0	Complete
3.A.1.120.1	(Putative) Drug Resistance ATPase-1 (Drug RA1)	1	ebA7225	SrmB	640	550	1.00E-56	0	Complete
3.A.1.120.3	(Putative) Drug Resistance ATPase-1 (Drug RA1)	1	ebA2876	OleB	612	569	2.00E-42	0	Complete
3.A.1.120.4	(Putative) Drug Resistance ATPase-1 (Drug RA1)	1	ebA6102	CarA	554	551	1.00E-47	0	Complete
3.A.1.121.2	(Putative) Drug Resistance ATPase-2 (Drug RA2)	1	ebA2792	VgrB	545	552	1.00E-52	0	Complete
3.A.1.122.1	Macrolide Exporter (MacB)	2	ebA1848	MacB	662	648	e-142	4	Complete
3.A.1.122.1	Macrolide Exporter (MacB)		ebA1846	MacA	448	371	1.00E-52	1	Complete
3.A.1.122.1	Macrolide Exporter (MacB)		ebA4059	MacB	402	648	1.00E-35	4	Probably complete
3.A.1.122.1	Macrolide Exporter (MacB)		ebA4061	SalX domain	232	648	1.00E-45	0 (TC 4)	COG0178,
3.A.1.122.1	Macrolide Exporter (MacB)	C(2)	ebA2970	MacB?	1888	648	3.00E-08	0 (TC 4)	UvrA, exonuclease ATPase subunit
3.A.1.123.1	Peptide-4 Exporter (Pep4E)	1	ebA2229	PepT	575	571	3.00E-49	6 (TC 5)	Complete
3.A.1.125.1	Lipoprotein Translocase (LPT)	1	ebA5819	LoIE (M)	417	413	3.00E-63	4	Complete
3.A.1.125.1	Lipoprotein Translocase (LPT)		ebA5820	LoID (C)	228	233	2.00E-66	0 (TC 1)	
3.A.1.125.1	Lipoprotein Translocase (LPT)		ebA4058	LoIC (M)	400	233	1.00E-33	4	
3.A.1.125.1	Lipoprotein Translocase (LPT)	1	ebA5805	LoIE (M)	849	413	3.00E-07	11 (TC 4)	LoIC missing (reverse blast result—LoIE)
3.A.1.125.1	Lipoprotein Translocase (LPT)		ebA5806	LoID (C)	239	233	8.00E-39	0	
3.A.1.125.1	Lipoprotein Translocase (LPT)		ebA6813	LoID (C)	231	233	2.00E-41	1 (TC 0)	?
3.A.1.125.1	Multidrug Resistance Exporter (MDR) (ABC _B)	1	ebA5186	BSEP	572	1321	7.00E-31	6 (TC 12)	Probably complete
3.A.1.210.3	Heavy Metal Transporter (HMT) (ABC _B)	2	ebA2601	ABC transporter homologue	604	609	c-154	6 (TC 8)	Complete
3.A.1.210.3	Heavy Metal Transporter (HMT) (ABC _B)		ebA6882	ABC transporter homologue	906	609	e-130	6 (TC 8)	Complete
3.A.1.212.1	Mitochondrial Peptide Exporter (MPE) (ABC _B)	1	ebA6704	MdIIp	600	695	4.00E-69	6 (TC 8)	Complete

Table 2 continued

TC subject protein	Family name	No. of complete systems	EbN1ID no.	TC homologue (protein name)	EbN1 query length	TC subject length	E-value	No. of TMSs	Estimated transporter status
3.A.2.1	H ⁺ - or Na ⁺ -translocating F-type, V-type, & A-type ATPase (F-ATPase) superfamily	1	ebA2999	ATP6	276	271	6.00E-61	6	Complete
3.A.2.1.1	H ⁺ - or Na ⁺ -translocating F-type, V-type, & A-type ATPase (F-ATPase) superfamily		ebA3000	ATPL	82	79	6.00E-08	2	
3.A.2.1.1	H ⁺ - or Na ⁺ -translocating F-type, V-type, & A-type ATPase (F-ATPase) superfamily		ebA3002	ATPF	157	156	5.00E-17	1	
3.A.2.1.1	H ⁺ - or Na ⁺ -translocating F-type, V-type, & A-type ATPase (F-ATPase) superfamily		ebA3003	ATPD	178	177	7.00E-24	0	
3.A.2.1.1	H ⁺ - or Na ⁺ -translocating F-type, V-type, & A-type ATPase (F-ATPase) superfamily		ebA3004	ATPA	512	513	0	0	
3.A.2.1.1	H ⁺ - or Na ⁺ -translocating F-type, V-type, & A-type ATPase (F-ATPase) superfamily		ebA3006	ATPG	289	287	1.00E-88	0	
3.A.2.1.1	H ⁺ - or Na ⁺ -translocating F-type, V-type, & A-type ATPase (F-ATPase) superfamily		ebA3007	ATPB	466	459	0	1	
3.A.2.1.1	H ⁺ - or Na ⁺ -translocating F-type, V-type, & A-type ATPase (F-ATPase) superfamily		ebA3008	ATPE	141	138	7.00E-36	0	
3.A.2.1.1	H ⁺ - or Na ⁺ -translocating F-type, V-type, & A-type ATPase (F-ATPase) superfamily		ebA224	Putative Ca2+-ATPase	911	905	0	10	Complete
3.A.3.2.4	P-type ATPase (P-ATPase) superfamily	2	ebA4146	Putative Ca2+-ATPase	897	905	e-143	10	Complete
3.A.3.2.4	P-type ATPase (P-ATPase) superfamily		ebA5128	CopA	817	727	1.00E-97	9 (TC 8)	Complete
3.A.3.5.1	P-type ATPase (P-ATPase) superfamily	2	ebA5154	CopA	803	727	e-141	8	Complete
3.A.3.5.1	P-type ATPase (P-ATPase) superfamily		ebA2180	Ag+-ATPase, SilP	785	824	0	8 (TC 9)	Complete
3.A.3.5.4	P-type ATPase (P-ATPase) superfamily	1	ebA699	CadA	694	727	1.00E-68	9	Complete
3.A.3.6.1	P-type ATPase (P-ATPase) superfamily	1	ebA1390	FtsE	220	222	3.00E-61	0 (TC 1)	Complete
3.A.5.1.1	General Secretory Pathway (Sec)	1	ebA1393	FtsY	386	497	4.00E-88	0	
3.A.5.1.1	General Secretory Pathway (Sec)		ebA1433	SecA	907	901	0	0 (TC 1)	
3.A.5.1.1	General Secretory Pathway (Sec)		ebA1417	SecD	635	615		5 (TC 6)	
3.A.5.1.1	General Secretory Pathway (Sec)		ebA1418	SecF	310	323		6	
3.A.5.1.1	General Secretory Pathway (Sec)		ebB40	YajC	108	110	1.00E-14	1	
3.A.5.1.1	General Secretory Pathway (Sec)		ebB90	RNPA	119	119	1.00E-06		
3.A.5.1.1	General Secretory Pathway (Sec)		ebC15	SecG	119	110	5.00E-04	3	
3.A.5.1.1	General Secretory Pathway (Sec)		ebA3810	SecE	115	127	3.00E-07	3	
3.A.5.1.1	General Secretory Pathway (Sec)		ebA3846	SecY	441	443	e-164	10	

Table 2 continued

TC subject protein	Family name	No. of complete systems	EbN1 ID no.	TC homologue (protein name)	EbN1 query length	TC subject length	E-value	No. of TMSS	Estimated transporter status
3.A.5.1.1	General Secretory Pathway (Sec)		ebA4351	Ffh	452	453	e-151	0	
3.A.7.4.1	Type IV (Conjugial DNA-Protein Transfer or VirB) Secretory Pathway (IVSP)		p2B10	TrbK	76	75	2.00E-09	0	Complete
3.A.7.4.1	Type IV (Conjugial DNA-Protein Transfer or VirB) Secretory Pathway (IVSP)		p2B2	TrbB	319	320	e-152	1	
3.A.7.4.1	Type IV (Conjugial DNA-Protein Transfer or VirB) Secretory Pathway (IVSP)		p2B3	TrbC	141	154	8.00E-35	3	
3.A.7.4.1	Type IV (Conjugial DNA-Protein Transfer or VirB) Secretory Pathway (IVSP)		p2B4	TrbE	845	852	0	2	
3.A.7.4.1	Type IV (Conjugial DNA-Protein Transfer or VirB) Secretory Pathway (IVSP)		p2B5	TrbF	261	260	4.00E-99	1	
3.A.7.4.1	Type IV (Conjugial DNA-Protein Transfer or VirB) Secretory Pathway (IVSP)		p2B6	TrbG	298	306	c-136	1	
3.A.7.4.1	Type IV (Conjugial DNA-Protein Transfer or VirB) Secretory Pathway (IVSP)		p2B8	TrbI	462	473	e-178	2	
3.A.7.4.1	Type IV (Conjugial DNA-Protein Transfer or VirB) Secretory Pathway (IVSP)		p2B9	TrbJ	252	254	9.00E-74	1	
3.A.7.4.1	Type IV (Conjugial DNA-Protein Transfer or VirB) Secretory Pathway (IVSP)		p2D1	TrbD	103	103	1.00E-50	2	
3.A.7.4.1	Type IV (Conjugial DNA-Protein Transfer or VirB) Secretory Pathway (IVSP)		p2B11	TrbL	548	572	e-113	9 (TC 8)	
3.A.11.1.1	Bacterial Competence-related DNA Transformation Transporter (DNA-T)	1	ebA5822	ComEC	782	776	2.00E-24	11 (TC 12)	Complete
3.A.11.1.1	Bacterial Competence-related DNA Transformation Transporter (DNA-T)		ebA3678	ComEA	155	205	1.00E-12	0 (TC 1)	
3.A.11.1.1	Bacterial Competence-related DNA Transformation Transporter (DNA-T)		ebA6555	ComFA	1157	463	1.00E-05	0	
3.A.12.1.1	Septal DNA Translocator (S-DNA-T)	1	ebA7041	SpoIIIIE	767	787	e-121	4	Complete
3.A.15.1.1	Outer Membrane Protein Secreting Main Terminal Branch (MTB)	1	ebA4900	PulL	405	398	2.00E-08	0	Probably complete
3.A.15.1.1	Outer Membrane Protein Secreting Main Terminal Branch (MTB)		ebA4904	PulK	307	326	2.00E-16	1	
3.A.15.1.1	Outer Membrane Protein Secreting Main Terminal Branch (MTB)		ebA4906	PulJ	184	198	2.00E-07	1	
3.A.15.1.1	Outer Membrane Protein Secreting Main Terminal Branch (MTB)		ebA4908	PulI	122	121	2.00E-12	1	
3.A.15.1.1	Outer Membrane Protein Secreting Main Terminal Branch (MTB)		ebA4910	?	176	140	0.41	1	
3.A.15.1.1	Outer Membrane Protein Secreting Main Terminal Branch (MTB)		ebA4913	PulF	400	381	2.00E-49	3	
3.A.15.1.1	Outer Membrane Protein Secreting Main Terminal Branch (MTB)		ebB170	PulG	147	140	3.00E-25	1	
3.A.15.1.1	Outer Membrane Protein Secreting Main Terminal Branch (MTB)		ebA4914	PulE	489	497	e-140	0	

Table 2 continued

TC subject protein	Family name	No. of complete systems	EbN1ID no.	TC homologue (protein name)	EbN1 query length	TC subject length	E-value	No. of TMSs	Estimated transporter status
3.A.15.1.1	Outer Bacterial Membrane Secretin (Secretin)		ebA4915	PilD	719	660	1.00E-72	1	
3.A.15.2.1	Outer Membrane Protein Secreting Main Terminal Branch (MTB)	1	ebA1770	PilT	347	344	e-148	0	Complete
3.A.15.2.1	Outer Membrane Protein Secreting Main Terminal Branch (MTB)		ebA1772	PilU	377	382	e-130	1	
3.A.15.2.1	Outer Membrane Protein Secreting Main Terminal Branch (MTB)		ebA1997	PilA	138	149	9.00E-06	2 (TC 1)	
3.A.15.2.1	Outer Membrane Protein Secreting Main Terminal Branch (MTB)		ebA4343	PilD	283	290	2.00E-75	8 (TC 6)	
3.A.15.2.1	Outer Membrane Protein Secreting Main Terminal Branch (MTB)		ebA4345	PilC	411	374	2.00E-95	4	
3.A.15.2.1	Outer Membrane Protein Secreting Main Terminal Branch (MTB)		ebA4346	PilB	571	566	e-173	1	
3.A.15.2.1	Outer Membrane Protein Secreting Main Terminal Branch (MTB)		ebA5112	FimT	173	168	6.00E-04	2 (TC 1)	
3.A.15.2.1	Outer Membrane Protein Secreting Main Terminal Branch (MTB)		ebD88	PilE	137	141	2.00E-07	2 (TC 1)	
3.A.15.2.1	Outer Membrane Protein Secreting Main Terminal Branch (MTB)		ebA2259	PilQ	721	714	e-130	1	
3.A.15.2.1	Outer Membrane Protein Secreting Main Terminal Branch (MTB)		ebA5118	FimU	184	168	7.00E-06	1 (TC 1)	
3.A. total		57							
3.D.1.2.1	Proton or Sodium Ion-translocating NADH Dehydrogenase (NDH)	1	ebB168	NQOB	101	101	3.00E-36	3	Complete
3.D.1.2.1	Proton or Sodium Ion-translocating NADH Dehydrogenases (NDH)		ebD11	NQO7	124	121	2.00E-34	3	
3.D.1.2.1	Proton or Sodium Ion-translocating NADH Dehydrogenase (NDH)		ebA4835	Nqo6	158	173	6.00E-53	1 (TC 0)	
3.D.1.2.1	Proton or Sodium Ion-translocating NADH Dehydrogenase (NDH)		ebA4836	NQO5	201	206	7.00E-37	1	
3.D.1.2.1	Proton or Sodium Ion-translocating NADH Dehydrogenase (NDH)		ebA4837	NQO4	417	412	e-148	0	
3.D.1.2.1	Proton or Sodium Ion-translocating NADH Dehydrogenase (NDH)		ebA4838	NQO2	159	239	2.00E-21	1	
3.D.1.2.1	Proton or Sodium Ion-translocating NADH Dehydrogenase (NDH)		ebA4840	NQO1	437	431	e-109	0 (TC 1)	
3.D.1.2.1	Proton or Sodium Ion-translocating NADH Dehydrogenase (NDH)		ebA4841	NQO3	779	672	5.00E-95	1	
3.D.1.2.1	Proton or Sodium Ion-translocating NADH Dehydrogenase (NDH)		ebA4842	NQO8	349	345	3.00E-91	8	
3.D.1.2.1	Proton or Sodium Ion-translocating NADH Dehydrogenase (NDH)		ebA4843	NQO9	161	163	1.00E-54	0 (TC 1)	

Table 2 continued

TC subject protein	Family name	No. of complete systems	EbN1ID no.	TC homologue (protein name)	EbN1 query length	TC subject length	E-value	No. of TMSs	Estimated transporter status
3.D.1.2.1	Proton or Sodium Ion-translocating NADH Dehydrogenases (NDH)	ebA4844	NQOA	200	200	2,00E-22	5		
3.D.1.2.1	Proton or Sodium Ion-translocating NADH Dehydrogenases (NDH)	ebA4846	NQOC	673	703	e-133	16		
3.D.1.2.1	Proton or Sodium Ion-translocating NADH Dehydrogenases (NDH)	ebA4847	NQOD	493	513	e-131	14		
3.D.1.2.1	Proton or Sodium Ion-translocating NADH Dehydrogenases (NDH)	ebA4848	NQOE	489	499	1,00E-59	14 (TC 15)		
3.D.2.2.1	Proton-translocating Transhydrogenase (PTH)	ebA4497	PNTAA	374	384	3,00E-75	0		Complete
3.D.2.2.1	Proton-translocating Transhydrogenase (PTH)	ebA4498	PNTAB	100	139	1,00E-10	3		
3.D.2.2.1	Proton-translocating Transhydrogenase (PTH)	ebA4500	PNTB	457	464	8,00E-99	10		
3.D.3.1.1	Proton-translocating Quinol:Cytochrome c Reductase (QCR) superfamily	ebA1197	CYB	440	444	5,00E-96	11 (TC 10)		Complete
3.D.3.1.1	Proton-translocating Quinol:Cytochrome c Reductase (QCR) superfamily	ebA1196	CY1	252	309	4,00E-14	2		
3.D.3.1.1	Proton-translocating Quinol:Cytochrome c Reductase (QCR) superfamily	ebA1198	UCRI	198	190	1,00E-23	1		
3.D.4.3.1	Proton-translocating Cytochrome Oxidase (COX) superfamily	ebA5131	Cox1	474	593	5,00E-10	12		Complete
3.D.4.3.1	Proton-translocating Cytochrome Oxidase (COX) superfamily	ebA4638	Cox2	336	331	3E-24	8 (TC 9)		
3.D.4.4.2	Proton-translocating Cytochrome Oxidase C(4) (COX) superfamily	ebA182	CtaE	199	205	4,00E-13	5		Incomplete
3.D.4.4.1	Proton-translocating Cytochrome Oxidase C(6) (COX) superfamily	ebA2294	COXX	295	305	5,00E-09	9		Incomplete
3.D.4.4.1	ebD10 membrane protein, putative accessory subunit of cytochrome c oxidase	ebD10	COXX	100					Complete
3.D.4.4.1	Proton-translocating Cytochrome Oxidase (COX) superfamily	ebA544	COX1	607	622	e-112	10 (TC 14)		
3.D.4.4.1	Proton-translocating Cytochrome Oxidase (COX) superfamily	ebA4547	COX2	310	356	5,00E-16	2 (TC 3)		
3.D.4.4.1	Proton-translocating Cytochrome Oxidase (COX) superfamily	ebA542	COX3						
3.D.4.4.1	Proton-translocating Cytochrome Oxidase (COX) superfamily	ebA4540	COX4	120	356	6,00E-05	0 (TC 3)		
3.D.4.4.2	Proton-translocating Cytochrome Oxidase C(4) (COX) superfamily	ebA6329	CtaE	198		2E-13	5		Incomplete
3.D.4.5.1	Proton-translocating Cytochrome Oxidase C(5) (COX) superfamily	ebA4237	CyoE	305	296	2,00E-30	9		Incomplete

Table 2 continued

TC subject protein	Family name	No. of complete systems	EbN1 ID no.	TC homologue (protein name)	EbN1 query length	TC subject length	E-value	No. of TMSS	Estimated transporter status
3.D.4.6.1	Proton-translocating Cytochrome Oxidase (COX) superfamily	1	ebA3665	CycA	177	155		2 (TC 3)	Probably complete
3.D.4.6.1	Proton-translocating Cytochrome Oxidase (COX) superfamily		ebA3666	CX1B	495	558	1.00E-15	12	
3.D.4.7.1	Proton-translocating Cytochrome Oxidase (COX) superfamily	1	ebA156	COX1 + COX3	836	622	e-136	18 (TC 13)	Complete
3.D.4.7.1	Proton-translocating Cytochrome Oxidase (COX) superfamily		ebA158	COX2	280	356	5.00E-16	2 (TC 3)	
3.D.4.8.1	Proton-translocating Cytochrome Oxidase (COX) superfamily	1	ebA4231	Cox3p					Probably complete
3.D.4.8.1	Proton-translocating Cytochrome Oxidase (COX) superfamily		ebA4227	Cox2p	301	227	7.00E-34	3 (TC 2)	
3.D.4.8.1	Proton-translocating Cytochrome Oxidase (COX) superfamily		ebA4228	Cox1p	526	514	e-149	12 (TC 13)	
3.D.4.8.1	Proton-translocating Cytochrome Oxidase (COX) superfamily		ebA4229	Cox1Ip					
3.D.4.8.1	Proton-translocating Cytochrome Oxidase (COX) superfamily		ebA4233	Shy1p					
3.D.4.10.1	Proton-translocating Cytochrome Oxidase (COX) superfamily	1	ebB6	NorC	198	261	1.00E-34	0 (TC 1)	Complete
3.D.4.10.1	Proton-translocating Cytochrome Oxidase (COX) superfamily		ebA179	NorB	460	593	1.00E-154	12	
3.D.6.1.1	Putative Ion (H^+ or Na^+)-translocating NADH:Ferredoxin Oxidoreductase (NFO)	2	ebA2575	RnfA	193	193	3.00E-59	6	Complete
3.D.6.1.1	Putative Ion (H^+ or Na^+)-translocating NADH:Ferredoxin Oxidoreductase (NFO)		ebA2576	RnfB	176	187	7.00E-29	1	
3.D.6.1.1	Putative Ion (H^+ or Na^+)-translocating NADH:Ferredoxin Oxidoreductase (NFO)		ebA2578	RnfD	362	358	1.00E-48	6 (TC 7)	
3.D.6.1.1	Putative Ion (H^+ or Na^+)-translocating NADH:Ferredoxin Oxidoreductase (NFO)		ebA2579	RnfC	508	519	e-117	0 (TC 2)	
3.D.6.1.1	Putative Ion (H^+ or Na^+)-translocating NADH:Ferredoxin Oxidoreductase (NFO)		ebA2581	RnfG	211	217	2.00E-44	1	
3.D.6.1.1	Putative Ion (H^+ or Na^+)-translocating NADH:Ferredoxin Oxidoreductase (NFO)		ebA2585	RnfE	225	441	1.00E-63	5 (TC 6)	
3.D.6.1.1	Putative Ion (H^+ or Na^+)-translocating NADH:Ferredoxin Oxidoreductase (NFO)		ebA2586	RnfH	101	85	3.00E-22	0	

Table 2 continued

TC subject protein	Family name	No. of complete systems	EbN1ID no.	TC homologue (protein name)	EbN1 query length	TC subject length	E-value	No. of TMSS	Estimated transporter status
3.D.6.1.1	Putative Ion (H^+ or Na^+)-translocating NADH:ferredoxin Oxidoreductase (NFO)		ebA4878	RnfG	232	441	2.00E-48	5 (TC 6)	Complete
3.D.6.1.1	Putative Ion (H^+ or Na^+)-translocating NADH:ferredoxin Oxidoreductase (NFO)		ebA4881	RnfG	220	217	3.00E-25	1	
3.D.6.1.1	Putative Ion (H^+ or Na^+)-translocating NADH:ferredoxin Oxidoreductase (NFO)		ebA4882	RnfD	335	358	2.00E-42	10 (TC 7)	
3.D.6.1.1	Putative Ion (H^+ or Na^+)-translocating NADH:ferredoxin Oxidoreductase (NFO)		ebA4883	RnfC	547	519	7.00E-87	2	
3.D.6.1.1	Putative Ion (H^+ or Na^+)-translocating NADH:ferredoxin Oxidoreductase (NFO)		ebA4884	RnfB	183	187	2.00E-37	2 (TC 1)	
3.D.6.1.1	Putative Ion (H^+ or Na^+)-translocating NADH:ferredoxin Oxidoreductase (NFO)		ebA4887	RnfA	194	193	1.00E-46	6	
3.D.6.1.1	Putative Ion (H^+ or Na^+)-translocating NADH:ferredoxin Oxidoreductase (NFO)		ebA6652	RnfH	107	85	7.00E-11	0	
3.D total		11							
4.A.2.1.1	PTS Fructose-Mannitol (Fru)	1	ebA3391	PTFA (IIA)	162	376	3.00E-06	0	Complete
4.A.2.1.1	PTS Fructose-Mannitol (Fru)		ebA1726	PTFB (IIB)	274	563	4.00E-04	8	
4.A.6.1.1	PTS Mannose-Fructose-Sorbitose (Man)		ebA2794	MANX (IIA)	128	322	6.00E-10	0	Incomplete
4.A total :									
5.A.2.1.1	Disulfide Bond Oxidoreductase-B (DsbB)	1	ebA3876	DsbB	166	176	1.00E-11	5 (TC4)	Complete
5.A.3.1.1	Prokaryotic Molybdopterin-containing Oxidoreductase (PMO)	1	ebA6282	Narl	232	225	4.00E-53	5	Complete
5.A.3.1.1	Prokaryotic Molybdopterin-containing Oxidoreductase (PMO)		ebA6286	NarG	1251	1246	0	0	
5.A.3.1.1	Prokaryotic Molybdopterin-containing Oxidoreductase (PMO)		ebA6285	NarH	516	514	0	0	
5.A.3.2.1	Disulfide Bond Oxidoreductase D (DsbD)	1	ebA2749	FdhG	951	1015	1.00E-35	2	Probably complete
5.A.3.2.1	Disulfide Bond Oxidoreductase D (DsbD)		ebA2750	FdhH + FdhI	585	294		4	
5.A.3.2.1	Prokaryotic Molybdopterin-containing Oxidoreductase (PMO)		ebA2935	FdnH	201	192	1.00E-26	0	Complete
5.A.3.2.1	Prokaryotic Molybdopterin-containing Oxidoreductase (PMO)		ebA2933	FdhI	375	217	2.00E-17	6 (TC 4)	
5.A.3.2.1	Prokaryotic Molybdopterin-containing Oxidoreductase (PMO)		ebA2936	FdnG	969	1015	2.00E-60	2	

Table 2 continued

TC subject protein	Family name	No. of complete systems	EbN1 ID no.	TC homologe (protein name)	EbN1 query length	TC subject length	E-value	No. of TMSs	Estimated transporter status
5.A.3.3.2	Prokaryotic Molybdopterin-containing Oxidoreductase (PMO)	1	ebA3328	Anaerobic dimethyl sulfide reductase chain A precursor a.k.a. DMSO reductase (DmsA)	694	814	6.00E-62	1	Complete
5.A.3.3.2	Prokaryotic Molybdopterin-containing Oxidoreductase (PMO)		ebA5004	Sulfoxide reductase chain B a.k.a. DMSO reductase iron-sulfur subunit (DmsB)	158	204	4.00E-17	0	
5.A.3.3.2	Prokaryotic Molybdopterin-containing Oxidoreductase (PMO)		ebA5396	Anaerobic dimethyl sulfide reductase chain C a.k.a. DMSO reductase anchor subunit (DmsC)	303	287	0.001	8	
5.A.3.4.1	Prokaryotic Molybdopterin-containing Oxidoreductase (PMO)	1	ebA1211	Trimethylamine oxidase I	939	848	4.00E-16	1 (TC 0)	Complete
5.A.3.4.1	Prokaryotic Molybdopterin-containing Oxidoreductase (PMO)		ebA1009	Cytochrome c-type protein torC	203	390	2.00E-09	2 (TC 1)	
5.A.3.4.1	Prokaryotic Molybdopterin-containing Oxidoreductase (PMO)		ebA5795	Chaperone protein torD	219	199	5.00E-04	0	
5.A.3.5.2	Prokaryotic Molybdopterin-containing Oxidoreductase (PMO)	1	ebA1012	Reducatase chain B a.k.a. sulfur reductase chain B	256	191	4.00E-38	1 (TC 0)	Complete
5.A.3.5.2	Prokaryotic Molybdopterin-containing Oxidoreductase (PMO)		ebA1013	Reducatase chain C a.k.a. sulfur reductase chain C	312	317	5.00E-16	8	
5.A.3.5.2	Prokaryotic Molybdopterin-containing Oxidoreductase (PMO)		ebA1017	Reducatase chain A precursor	841	763	1.00E-45	0	
5.A.3.5.2	Prokaryotic Molybdopterin-containing Oxidoreductase (PMO)	1	ebA6426	Reducatase chain A precursor a.k.a. sulfur reductase chain A	877	763	3.00E-63	0	Complete
5.A.3.5.2	Prokaryotic Molybdopterin-containing Oxidoreductase (PMO)		ebA6432	Reducatase chain C a.k.a. sulfur reductase chain C	283	317	8.00E-08	8	
5.A.3.5.2	Prokaryotic Molybdopterin-containing Oxidoreductase (PMO)		ebA6435	Reducatase chain B a.k.a. sulfur reductase chain B	236	257	2.00E-50	0	
5.A.3.9.1	Prokaryotic Molybdopterin-containing Oxidoreductase (PMO)	2	c1A62	Anaerobic ethylbenzene dehydrogenase subunit C	214	214	e-122	0	Complete

Table 2 continued

TC subject protein	Family name	No. of complete systems	EbN1ID no.	TC homologue (protein name)	EbN1 query length	TC subject length	E-value	No. of TMSs	Estimated transporter status
5.A.3.9.1	Prokaryotic Molybdopterin-containing Oxidoreductase (PMO)		c1A63	Anaerobic ethylbenzene dehydrogenase subunit B	352	352	0	0	
5.A.3.9.1	Prokaryotic Molybdopterin-containing Oxidoreductase (PMO)		c1A65	Ethylbenzene dehydrogenase subunit A	974	974	0	1	Complete
5.A.3.9.1	Prokaryotic Molybdopterin-containing Oxidoreductase (PMO)		ebA5790	Ethybenzene dehydrogenase subunit A	974	974	0	1	Complete
5.A.3.9.1	Prokaryotic Molybdopterin-containing Oxidoreductase (PMO)		ebA5791	Anaerobic ethylbenzene dehydrogenase subunit B	352	352	0	0	
5.A.3.9.1	Prokaryotic Molybdopterin-containing Oxidoreductase (PMO)		ebA5793	Anaerobic ethylbenzene dehydrogenase subunit C	214	214	8.00E-71	0	
5.A.3.10.1	Prokaryotic Molybdopterin-containing Oxidoreductase (PMO)	2	ebA5622	Tetrathionate reductase subunit A	868	1053	2.00E-28	3 (TC 0)	Complete
5.A.3.10.1	Prokaryotic Molybdopterin-containing Oxidoreductase (PMO)		ebA5630	Tetrathionate reductase subunit B	200	257	7.00E-42	0	
5.A.3.10.1	Prokaryotic Molybdopterin-containing Oxidoreductase (PMO)		ebA5631	Tetrathionate reductase subunit C	308	344	6.00E-07	8 (TC 9)	
5.A.3.10.1	Prokaryotic Molybdopterin-containing Oxidoreductase (PMO)		ebA3055	Tetrathionate reductase subunit B	247	257	8.00E-69	1 (TC 0)	
5.A.3.10.1	Prokaryotic Molybdopterin-containing Oxidoreductase (PMO)		ebA3056	Tetrathionate reductase subunit C	362	344	7.00E-45	9	Complete
5.A.3.10.1	Prokaryotic Molybdopterin-containing Oxidoreductase (PMO)		ebD4	Tetrathionate reductase subunit A	1027	1053	0	1 (TC 0)	
5.A total		12							
5.B.1.6.1	gp91phox Phagocyte NADPH Oxidase-associated Cytochrome b558 (CytB) H ⁺ -channel	4	ebA3186	Putative oxido-reductase	439	450	5.00E-58	7	Complete
5.B.1.6.1	gp91phox Phagocyte NADPH Oxidase-associated Cytochrome b558 (CytB) H ⁺ -channel		ebA4977	Putative oxido-reductase	442	450	3.00E-80	7	Complete
5.B.1.6.1	gp91phox Phagocyte NADPH Oxidase-associated Cytochrome b558 (CytB) H ⁺ -channel		ebA7152	Putative oxido-reductase	439	450	9.00E-63	6 (TC 7)	Complete

Table 2 continued

TC subject protein	Family name	No. of complete systems	EbN1 ID no.	TC homologue (protein name)	EbN1 query length	TC subject length	E-value	No. of TMSS	Estimated transporter status
5.B.1.1	gp91phox Phagocyte NADPH Oxidase-associated Cytochrome b558 (CytB) H ⁺ -channel	4	ebA2594	Putative oxidoreductase	159	450	5.00E-12	3 (TC 7)	Complete
5.B. total			eba3446	EmrA	433	390	1.00E-78	2 (TC 1)	Complete
8.A.1.1.1	Membrane Fusion Protein (MFP)	1	eba6248	CzcB	383	395	7.00E-10	1	Complete
8.A.1.2.1	Membrane Fusion Protein (MFP)	1	eba1673	HlyD	457	478	1.00E-13	1	Complete
8.A.1.3.1	Membrane Fusion Protein (MFP)	3	eba1805	HlyD	473	478	6.00E-34	1	Complete
8.A.1.3.1	Membrane Fusion Protein (MFP)		eba7180	HlyD	434	478	4.00E-12	1	Complete
8.A.1.3.1	Membrane Fusion Protein (MFP)		eba3872	AcrA	349	397	1.00E-10	1	Complete
8.A.1.6.1	Membrane Fusion Protein (MFP)	3	eba4062	AcrA	390	397	1.00E-04	1	Complete
8.A.1.6.1	Membrane Fusion Protein (MFP)		eba7103	AcrA	386	397	9.00E-22	1	Complete
8.A.3.1.1	Cytoplasmic Membrane-Periplasmic Auxiliary-1 (MPA1) Protein with Cytoplasmic (C) Domain (MPA1-C or MPA1 + C)	1	eba6797	ExoP	760	786	5.00E-45	2 (TC 3)	Complete
8.A.5.1.3	K ⁺ Transport/Nucleotide-binding Regulatory Domain/Protein (KTN)	1	p2A379	β 1a	336	401	2.00E-19	0	Complete
8.A.7.1.1	Phosphotransferase System Enzyme I (EI)	1	eba2795	Enzyme I	576	575	5.00E-91	2 (TC 0)	Complete
8.A.7.1.1	Phosphotransferase System Enzyme I (EI)		eba5781	PEP synthetase	592	575	6.00E-06	0	Probably complete
8.A.7.1.1	Phosphotransferase System Enzyme I (EI)	1	eba5830	Phenyl-phosphate synthetase	788	575	1.00E-21	0	Complete
8.A.8.1.1	Phosphotransferase System HPr (HPr)	1	ebD68	HPr	89	85	1.00E-05	0	Complete
8.A.9.1.1	rBAT Transport Accessory Protein (rBAT)	3	eba583	rBAT	562	677	8.00E-65	1	Complete
8.A.9.1.1	rBAT Transport Accessory Protein (rBAT)		eba6923	rBAT	767	677	1.00E-04	0 (TC 1)	Complete
8.A.9.1.1	rBAT Transport Accessory Protein (rBAT)		eba7001	rBAT	1114	677	9.00E-57	1	Complete
8.A. total		16							
9.A.8.1.1	Ferrous Iron Uptake (FeoB)	2	eba503	FeoB (GTPase)	467	773	8.00E-04	1 (TC 11)	Probably complete
9.A.8.1.1	Ferrous Iron Uptake (FeoB)		eba5540	FeoB (GTPase)	305	773	9.00E-08	0 (TC 11)	Probably complete
9.A.8.1.3	Ferrous Iron Uptake (FeoB)	2	eba1256	FeoB	442	614	8.00E-09	1 (TC 12)	Probably complete
9.A.8.1.3	Ferrous Iron Uptake (FeoB)		eba4139	FeoB	363	614	1.00E-06	1 (TC 12)	Probably complete
9.A.19.1.1	Mg ²⁺ Transporter-E (MgeE)	1	eba1741	MgeE	522	312	3.00E-36	5	Complete
9.A.40.1.2	HlyC/CorC (HCC) of Putative Transporters	1	eba1335	CorC	282	292	3.00E-59	0 (TC 1)	Complete
9.A.40.2.1	HlyC/CorC (HCC) of Putative Transporters	2	eba4147	YrkA	444	434	2.00E-27	3 (TC 4)	Complete

Table 2 continued

TC subject protein	Family name	No. of complete systems	EbN1ID no.	TC homologue (protein name)	EbN1 query length	TC subject length	E-value	No. of TMSs	Estimated transporter status
9.A.40.2.1	HlyC/CorC (HCC) of Putative Transporters		ebA6189	YrkA	432	434	1.00E-33	4	Complete
9.A. total		8							
9.B.3.1.1	Putative Bacterial Murein Precursor Exporter (MPE)	1	ebA1445	FtsW	410	414	2.00E-67	10	Complete
9.B.3.1.2	Putative Bacterial Murein Precursor Exporter (MPE)	1	ebA3041	RodA	380	370	3.00E-73	10 (TC 9)	Complete
9.B.11.1.1	Putative ER-chloroplast Lipid Translocase (ECLT)	1	ebA1312	Tgdl protein	264	350	3.00E-23	6 (TC 5)	Complete
9.B.14.1.1	Putative Heme Exporter Protein (HEP)	2	ebA2222	CcmF (Ccl1)	713	653	1.00E-05	15	Complete
9.B.14.1.1	Putative Heme Exporter Protein (HEP)	1	ebA3521	CcmF (Ccl1)	657	653	e-168	15	Complete
9.B.17.1.4	Putative Fatty Acid Transporter (FAT)	15	ebA1206	FadD	563	561	2.00E-30	1 (TC 2)	Complete
9.B.17.1.4	Putative Fatty Acid Transporter (FAT)		ebA1960	FadD	549	561	3.00E-39	1 (TC 2)	Complete
9.B.17.1.4	Putative Fatty Acid Transporter (FAT)		ebA2757	FadD	534	561	7.00E-41	2	Complete
9.B.17.1.4	Putative Fatty Acid Transporter (FAT)		ebA321	FadD	664	561	2.00E-05	2	Complete
9.B.17.1.4	Putative Fatty Acid Transporter (FAT)		ebA4297	FadD	546	561	3.00E-43	1 (TC 2)	Complete
9.B.17.1.4	Putative Fatty Acid Transporter (FAT)		ebA4326	FadD	522	561	2.00E-20	1 (TC 2)	Complete
9.B.17.1.4	Putative Fatty Acid Transporter (FAT)		ebA4661	FadD	562	561	2.00E-67	0 (TC 2)	Complete
9.B.17.1.4	Putative Fatty Acid Transporter (FAT)		ebA4749	FadD	555	561	0	3 (TC 2)	Complete
9.B.17.1.4	Putative Fatty Acid Transporter (FAT)		ebA5317	FadD	624	561	2.00E-12	0 (TC 2)	Complete
9.B.17.1.4	Putative Fatty Acid Transporter (FAT)		ebA5705	FadD	831	561	4.00E-27	4 (TC 2)	Complete
9.B.17.1.4	Putative Fatty Acid Transporter (FAT)		ebA6518	FadD	550	561	4.00E-37	0 (TC 2)	Complete
9.B.17.1.4	Putative Fatty Acid Transporter (FAT)		ebA727	FadD	523	561	1.00E-46	2	Complete
9.B.17.1.4	Putative Fatty Acid Transporter (FAT)		ebA5301	FadD	533	561	3.00E-48	1 (TC 2)	Complete
9.B.17.1.4	Putative Fatty Acid Transporter (FAT)		ebA5368	FadD	493	561	2.00E-24	2	Complete
9.B.17.1.4	Putative Fatty Acid Transporter (FAT)		ebA172	FadD	662	561	4.00E-26	2	Complete
9.B.17.1.6	Putative Fatty Acid Transporter (FAT)	6	ebA2050	CaiC	510	522	1.00E-46	1 (TC 0)	Complete
9.B.17.1.6	Putative Fatty Acid Transporter (FAT)		ebA3433	CaiC	546	522	4.00E-38	0	Complete
9.B.17.1.6	Putative Fatty Acid Transporter (FAT)		ebA7220	CaiC	630	522	6.00E-12	3 (TC 0)	Complete
9.B.17.1.6	Putative Fatty Acid Transporter (FAT)		p2A386	CaiC	558	522	8.00E-64	3	Complete
9.B.17.1.6	Putative Fatty Acid Transporter (FAT)		eba4666	CaiC	662	522	5.00E-08	1 (TC 0)	Complete
9.B.17.1.6	Putative Fatty Acid Transporter (FAT)		ebA4717	CaiC	663	522	1.00E-25	0	Complete
9.B.20.2.1	Putative Mg ²⁺ Transporter-C (MgtC)	1	ebB238	SrpB	159	182	9.00E-27	5	Complete
9.B.24.2.1	Testis-Enhanced Gene Transfer (TEGT)	2	ebA4143	YccA	221	219	4.00E-58	7	Complete
9.B.24.2.1	Testis-Enhanced Gene Transfer (TEGT)		p1B299	YccA	220	219	2.00E-55	7	Complete
9.B.26.1.1	PF27 (PF27)	1	ebA1709	Y615	192	206	9.00E-07	5 (TC 6)	Complete
9.B.27.1.1	Ydix-Z (Ydix-X-Z)	1	ebA6946	Ydix	743	236	3.00E-07	7 (TC 6)	Complete
9.B.31.1.1	YqfH (YqfH)	1	ebA4373	YgH	197	205	1.00E-28	5	Complete
9.B.32.1.2	Putative Vectorial Glycosyl Polymerization (VGP)	1	ebA6116	IcaA	905	412	2.00E-07	9 (TC 4)	Complete

Table 2 continued

TC subject protein	Family name	No. of complete systems	EbN1 IID no.	TC homologue (protein name)	EbN1 query length	TC subject length	E-value	No. of TMSSs	Estimated transporter status
9.B.32.1.3	Putative Vectorial Glycosyl Polymerization (VGP)	3	ebA1112	YcdQ	234	441	5.00E-04	0	Probably complete
9.B.32.1.3	Putative Vectorial Glycosyl Polymerization (VGP)		ebA600	YcdQ	247	441	8.00E-09	5	Complete
9.B.32.1.3	Putative Vectorial Glycosyl Polymerization (VGP)		ebA7076	YcdQ	287	441	1.00E-07	2 (TC 5)	Complete
9.B.42.1.1	ExeAB Secretin Assembly/Export Complex		ebA4256	ExeA	360	547	2.00E-50	1 (TC 2)	Incomplete
9.B.60.2.1	YfcA	1	ebA3571	DshD	628	601	2.00E-86	10 (TC 9)	Complete
9.B.67.1.1	Putative Inorganic Carbon (HCO ₃ ⁻) Transporter/O-Antigen Polymerase (ICT/OAP)	1	ebA4277	IctB	444	467	6.00E-04	10	Complete
9.B total		38							

closely related to the Neisserial Omp85 homologue (Wu et al. 2005). It also has a single YfiO homologue, a single NlpB homologue, and a single YfgL homologue. OmpIP complexes function to insert β -structured proteins into the outer membrane.

Finally, strain EbN1 has one homologue of the OstA protein, proposed to function in lipopolysaccharide export across the outer membrane (TC 1.B.42) (Bos et al. 2004; Hu and Saier 2006). It also has an RlpB homologue that in *E. coli* functions in conjunction with OstA (Wu et al. 2006).

Secondary Carriers

The largest superfamily of secondary carriers found in nature is the major facilitator superfamily (MFS) (Chang et al. 2004). Strain EbN1 has 22 (8.5% of the total transporters) recognized members of this superfamily, but surprisingly it contains no MFS sugar transporters. In fact, this organism has no primary or secondary transporters that take up sugars. A single fructose-type PTS permease (see below) is the only system that serves as a good candidate for sugar uptake. The deficiency in sugar uptake systems agrees well with the apparent inability of strain EbN1 to utilize sugars as growth substrates (Rabus and Widdel 1995).

Recognized MFS permeases in strain EbN1 include five putative drug (MDR):proton antiporters (four in family 2.A.1.2 and one in family 2.A.1.3). It also has two probable nitrate/nitrite antiporters and one nitrate:H⁺ symporter, all of family 2.A.1.8. There is a distant member of the oxalate/formate antiporter family (2.A.1.11) and seven systems for the potential uptake of aromatic compounds. These include four potential systems of TC family 2.A.1.15 (aromatic acid:H⁺ symporter), one of the phenyl propionate permease (PPP) family (2.A.1.27), one of the putative abietane diterpenoid transporter (ADT) family (TC 2.A.1.30), and one of the acriflavine sensitivity (YnfM) family (2.A.1.36.1; unpublished preliminary observations). Another MFS aromatic uptake permease may transport compounds resembling folate/biopterin (a member of TC family 2.A.71). Strain EbN1 may have a single cyanate uptake permease. It also has a single putative peptide uptake system of the PAT family (TC 2.A.1.25) and two homologues of the lysophospholipid uptake (LplT) family. LplT family proteins are required for periplasmic lipoprotein acylation as has been demonstrated in *E. coli* (Harvat et al. 2005). We also identified two MFS permeases of unknown function (see Table 2).

Strain EbN1 has two members of the CDF family of divalent cation diffusion facilitators. These systems are usually responsible for the active export of heavy metals such as Co²⁺, Cd²⁺, Zn²⁺, Ni²⁺, Hg²⁺, and Fe²⁺. Both of

the CDF systems in strain EbN1 are likely to be of broad specificity. Strain EbN1 also has a single heavy metal efflux pump of the RND superfamily (Table 2). This unusual system probably functions with auxiliary proteins to directly export heavy metals from the cytoplasm to the external milieu. Similarly to the CusABC system of *E. coli* (TC 2.A.6.1.4), it requires a periplasmic binding receptor. There are no other secondary carrier candidates for heavy metal efflux; only a single Fe^{3+} primary active uptake system of the ABC superfamily was identified. It appears that strain EbN1 takes up metals primarily via a CorA channel and expels them via CDF- and RND-type systems.

Within the RND superfamily (TC 2.A.6), we found 13 putative MDR efflux pumps of two different families (HAE1 [2.A.6.2] and HAE3 [2.A.6.7]), a single SecDF pair of auxiliary proteins TC 2.A.6.4.1 of the general secretory (Sec; TC 3.A.5.1.1) pathway, and a single member of the brominated aryl polyene pigment exporter family (2.A.6.8).

The drug/metabolite transporter (DMT) superfamily (Jack et al. 2001) is represented by one member of the four-TMS small multidrug resistance (SMR) family of cationic drug exporters and five members of the 10-TMS metabolite exporters of the DME family. Most of these proteins exhibit low BLAST scores (e^{-6} – e^{-10}), but our analyses clearly suggest that they are true members of the DME family within the DMT superfamily. Their substrate specificities cannot be reliably ascertained.

Another important superfamily for the export of macromolecules and drugs is the multidrug/oligosaccharidyl-lipid/polysaccharide (MOP) superfamily (TC 2.A.66) (Hvorup et al. 2003). Three of the twelve families within the MOP superfamily are represented in strain EbN1. One of these is likely to be specific for drugs; one undoubtedly exports polysaccharides; and one (within the mouse virulence factor [MVF] family) is of unknown specificity, as no member of this family has been functionally characterized.

Bacterial Oxa1 family members (YidC homologues; family 2.A.9) can function together with the general secretory (Sec) pathway to insert proteins into the cytoplasmic membrane. These proteins may also be able to insert some integral membrane proteins without the aid of the Sec system (Chen et al. 2002, 2003; Luirink et al. 2001; Yen et al. 2001). These YidC homologues form a complex with the SecDF proteins of the RND superfamily (Nouwen and Driessens 2002).

Strain EbN1 has 17 secondary carriers that are probably specific for various organic and inorganic anions in addition to those in the MFS. These fall into nine TC families: (1) one is in the tellurite/dicarboxylate transporter (TDT) family (TC 2.A.16); (2) one is in the phosphate transporter (PiT) family (TC 2.A.20); (3) two putative acetate/

glyoxylate uptake systems and a phenyl acetate uptake permease are in the SSS family (TC 2.A.21); (4) one putative sulfate transporter of the CHR family (TC 2.A.51) consists of two half-sized proteins as occurs in *B. subtilis* (Nies et al. 1998); (5) two possible sulfate transporters belong to the SulP family (2.A.53); (6) five complete systems are in the tripartite TRAP-T family, each with three components as expected (TC 2.A.56) (Kelly and Thomas 2001; Mulligan et al. 2007; Rabus et al. 1999), all of which may take up organic anions (see TCDB); (7) one member belongs to the arsenical resistance (Acr3) family (TC 2.A.59) of the BART superfamily (Mansour et al. 2007); (8) two possible malate/malonate transporters are in the AEC family (TC 2.A.69); and (9) a single member of the putative HCO_3^- transporter is in the ICT family (TC 9.B.69). There is some confusion in the literature concerning the ICT family. Members of this family may actually be o-antigen polymerases involved in lipopolysaccharide synthesis (see description in TC 9.B.69).

Strain EbN1 also has four recognizable primary active uptake transporters of the ABC superfamily specific for inorganic anions. These are probably specific for (1) sulfate (taken up by a member of the SulT family; TC 3.A.1.6) [the thiosulfate receptor that in *E. coli* can function with this transporter was not found]; (2) tungstate, taken up by a homologue of the TupABC system of *Eubacterium acidaminophilum*, also of the SulT family; (3) phosphate, taken up by a homologue of the *E. coli* high-affinity Pst porter of the PhoT family (3.A.1.7); and (4) molybdate, taken up by a homologue of the MolT family system of *E. coli*.

Amino acids, organic cations, and peptides are transported in strain EbN1 by at least six secondary porters and eight ABC-type active transporters, and these fall into six (super)families as follows: (1) peptides can probably be taken up via a member of the PAT family (TC 2.A.1.25) within the MFS as mentioned previously; (2) short semipolar amino acids such as alanine and glycine can be taken up via a member of the AGCS family (TC 2.A.25); (3) choline may be taken up via a member of the SSS family (TC 2.A.21.8), and interestingly, a member of the same family may be a sensor for amino acids, part of a sensor kinase (Nishijyo et al. 2001); (4) a single member of the TRAP-T family of tripartite systems may take up ectoine (a zwitterionic compound) and its derivatives (Grammann et al. 2002); and (5) finally, six ATP-hydrolysis-driven transport systems of the ABC superfamily are likely to take up amino acids. Interestingly, five of these are in the HAAT family (TC 3.A.1.4), and four of them probably transport hydrophobic (and sometimes semipolar) amino acids, while the fifth shows greatest similarity to a urea uptake porter of cyanobacteria (TC 3.A.1.4.4). There is only one member of the PAAT family (TC 3.A.1.3) which

undoubtedly takes up polar amino acids as is characteristic of the members of this family. This porter may take up acidic amino acids based on its best fit in TCDB (see Table 2). There are also two possible peptide uptake permeases of the PepT family (TC 3.A.1.5) in the ABC superfamily; one of these may have specificity for glutathione (Suzuki et al. 2005), which is important as a cytoplasmic reducing agent.

Monovalent inorganic cation transporters are numerous in strain EbN1, and these fall into five (super)families. (1) Two monovalent cation:proton antiporters are present. One belongs to the CPA1 family; it could be a Na^+/H^+ antiporter. The other belongs to the CPA2 family. Both of these families are within the CPA superfamily. The CPA2 family member could be a glutathione-regulated K^+ efflux porter. (2) Three members of the Trk family of heterodimeric K^+ uptake systems are present. (3) We found two members of the Amt family of probable NH_3 or NH_4^+ channel proteins (Fong et al. 2007; Ishikita and Knapp 2007; Javelle et al. 2005; Khademi et al. 2004). (4) One complete member of the multicomponent CPA3 family is present, with two of the seven constituents (the PhaA and PhaB equivalents) fused in a single large polypeptide chain. These complex permeases may be energized by NAD oxidoreduction. (5) Finally, there are two members of the KUP family of K^+ uptake permeases. Primary active monovalent cation transporters are described below.

Outer Membrane Transport Energizers

The TonB heteromeric complexes span the cytoplasmic membrane and the periplasm and interact with outer membrane receptors in Gram-negative bacteria. Homologues have been found only in Gram-negative bacteria. *E. coli* has two types of homologous outer membrane transport energizers, the TonB-ExbB-ExbD system and the TolA-TolQ-TolR system, both using the proton motive force and both in the TonB family (TC 2.C.1). These energizers act together with OMR-type outer membrane receptors. They have overlapping but distinct functions.

The TonB system of *E. coli* energize transport (uptake) via OMR-type porins (TC 1.B.14) specific for vitamin B^{12} , iron siderophores, group B colicins, and the DNA of filamentous bacteriophage such as $\phi 80$ and T1. They may also be involved in the extrusion of drugs and organic solvents. The TolA system transports group A colicins and the DNA of other filamentous phages. Colicin import requires close proximity of the inner and outer membranes. Loss of one of the TolA-TolQ-TolR proteins results in loss of periplasmic enzymes and increased sensitivity to drugs and bile salts. Surface localization of O-antigen lipopolysaccharide in *E. coli* depends on the TolA protein, possibly explaining the leakiness of TolA mutants. The *E. coli* TolA/Pal system

has also been reported to be necessary for the uptake of certain solutes (sugars, polyols, amino acids) (Bouveret et al. 2002; Cao and Klebba 2002; Llamas et al. 2003; Lloubes et al. 2001).

Different bacteria have different numbers and sets of these proteins. Strain EbN1 has two TonB-ExbBD systems, one TolAB-TolRQ-YbgF system, and eight OMR-type outer membrane receptors. Although we were able to identify all other constituents of this last system, a TolA-like protein, similar throughout its length to the *E. coli* TolA, could not be found. However, EbA2648 has a distant homologue of TonB, and this protein may serve the function of TolA. TonB and TolA of *E. coli* are distant homologues of each other.

Primary Active Transporters

The ABC Superfamily

Table 2 reports the ABC systems of strain EbN1 that resemble those in TCDB. In addition to the six amino acid uptake systems, the four inorganic anion uptake systems, and the two taurine uptake systems noted above, strain EbN1 has a single Fe^{3+} uptake transporter of the FeT family (TC 3.A.1.10), a $\text{Mn}^{2+}/\text{Zn}^{2+}/\text{Fe}^{2+}$ chelate uptake transporter of the MZT family (3.A.1.15.2), and a vitamin B_{12} uptake system (3.A.1.13). Surprisingly, strain EbN1 has no additional recognizable complete ABC uptake systems. It does, however, have many ABC-type multidrug efflux systems belonging to five different ABC families known to be involved in their extrusion. Most of these (11 systems including MsbA which can transport both lipids and drugs) (Reuter et al. 2003) may export hydrophobic and amphipathic substances including toxic metabolites, thereby protecting the organism. We also found a single putative heme exporter (TC 3.A.1.10.7), two probable heavy metal exporters similar to those in eukaryotes (TC 3.A.1.210), and two sequence dissimilar putative peptide exporters (TC 3.A.1.123 and 3.A.1.212), one resembling prokaryotic systems and one more similar to eukaryotic mitochondrial systems.

Ten macromolecular ABC-type efflux systems, three complete systems specific for lipooligosaccharides (3.A.1.102), one for lipopolysaccharides or their precursors (3.A.1.103), one specific for lipids (MsbA-type; 3.A.1.106), one specific for lipoproteins (LPT family; 3.A.1.125, as well as one incomplete system of this type), and three specific for proteins (TCs 3.A.1.109 and 3.A.1.110). All such systems use ATP hydrolysis to drive macromolecular export, and they are often coupled to membrane fusion proteins (MFP; 8.A.1) and outer membrane channel forming factors (OMF; 1.B.17) to pump solutes across both membranes and the periplasm of the Gram-negative bacterial envelope.

Other ATP-Driven Transporters

Strain EbN1 has a single F-type ATPase (3.A.2) for the interconversion of chemiosmotic (pmf) and chemical (ATP) energy. All eight expected constituents were identified (α , β , γ , δ , ε , a , b , c). It also has six full-length P-type ATPases, all probably specific for monovalent or divalent cations (one for Ag^+ , two for Cu^{+} , one for Cd^{2+} and other heavy metals, and two for Ca^{2+}). No other pyrophosphate hydrolysis-driven ion pumps were identified.

Protein Secretion Systems

Strain EbN1 has an ATP/GTP/pmf-driven general secretory (Sec) pathway, undoubtedly the most important system for protein secretion from the cytoplasm to the periplasm (Cao and Saier 2003). All of the components of the Sec system found in *E. coli* are also present in strain EbN1. Strain EbN1 has a three-component pmf-driven Tat system (TC 2.A.64), as do many bacteria (Yen et al. 2002b). The system in strain EbN1 has TatA, -C, and -E but seems to lack TatB. TatA, -B, and -E are homologous to each other, and only one of these homologues is absolutely required (Yen et al. 2002b). While the TatA, -B, and -E proteins form an expandable pore that accommodates fully folded and assembled substrate protein complexes, the TatC protein recognizes the substrate complex and nucleates pore formation (Holzapfel et al. 2007).

Three ABC-type I protein secretion systems were identified as discussed above. These systems transport their protein substrates across both membranes of the Gram-negative bacterial envelope in a single energy-coupled step. This process depends on a membrane fusion protein (MFP; TC 8.A.1) and an outer membrane factor (OMF; TC 1.B.17). Strain EbN1 has nine recognizable MFP homologues and six identifiable OMF homologues. An OMF-like TolC of *E. coli* can function with multiple transporter-MFP pairs (Yen et al. 2002a). The three ABC type I exporters most resemble characterized systems specific for (1) a hemolysin (HlyB), (2) hydrolytic enzymes such as proteases and lipase (LapB), and (3) an adhesin (HasD). It should be noted that MFPs and OMFs are very sequence divergent, so the possibility of additional homologues cannot be excluded.

Strain EbN1 has a single type IV (conjugal) DNA-protein secretory system (Cao and Saier 2001). The system most resembles the 10-component Trb system of plasmid IncP from *Enterobacter aerogenes*. All 10 proteins were identified, encoded on a strain EbN1 plasmid (Table 2). This *A. aromaticum* strain also has a three-component bacterial competence-related DNA transformation transporter (DNA-T) (Dubnau 1999; Kidane and Graumann 2005). This system resembles the *B. subtilis* DNA-T

system. A septal DNA translocator (S-DNA-T), resembling SpoIIIE, a well-characterized system in *B. subtilis*, is also present. Homologues of this protein are found in most, if not all, bacteria, where they function to translocate DNA across newly formed septa.

The *B. subtilis* DNA-T competence system has three components, Cme1 (ComEA), Cme3 (ComEC), and Cmf1 (ComEF). The Cmf1 homologue in strain EbN1 is nearly three times longer than its *B. subtilis* homologue. These two proteins show an overlap of about 300 residues, and this overlapping region includes two domains, the DEXH box helicases domain, including a Walker B motif for ATP binding, and the HELICc domain, also characteristic of helicases. The extra N-terminal 600 residues, lacking in the *B. subtilis* Cmf1 protein, includes a UvrB domain, a helicase subunit of the DNA excision repair complex involved in DNA replication, recombination and repair, and a CarD-TrcF domain, involved in binding to stalled RNA polymerase. At the C-terminus of this protein is a TrcF domain of unknown function. It seems likely that this large protein in strain EbN1 is multifunctional, possibly coordinating DNA uptake with recombination of the single DNA strand into the bacterial chromosome.

Strain EbN1 has many proteins showing homology to the Pul (pullulanase secretion system) and Pil (pilus secretion system), main terminal branch (MTB) protein secretion type II systems. Interestingly, 8 chromosomally encoded proteins most resemble Pul proteins (TC 3.A.15.1.1) of *Klebsiella pneumoniae*, and 10 chromosomally encoded proteins most resemble the homologous Pil proteins (TC 3.A.15.2.1) of *Pseudomonas aeruginosa*. Type II secretion systems do not all have the same numbers of constituents (Peabody et al. 2003; Sandkvist 2001). Thus, while the Pil-like system of strain EbN1 has all of the constituents recognized for the *P. aeruginosa* Pil system, the Pul-like system is not complete by the same criterion. We postulate that strain EbN1 has a chromosomally encoded MTB-type protein secretion system, a chromosomally encoded Pil-type pilus subunit secretion system, and a plasmid-encoded Trb system for protein/DNA export during conjugation.

Electron Flow-Driven H^+/Na^+ Pumps

Bacteria have a variety of energy-coupled electron flow-carrier complexes. Strain EbN1 has 15 components of a single Na^+ -translocating NADH dehydrogenase (NDH) complex and three constituents of a single complete proton-translocating transhydrogenase (PTH). Three components of a single proton-translocating quinol:cytochrome *c* reductase (QCR) are also present. Multiple subunits of as many as five distinct cytochrome oxidase (COX) systems were identified, but it is not certain that all of these are

complete. Several of these subunits most resemble subunits of the mammalian COX system (Table 2; see TCDB—www.tcdb.org). Interestingly, *E. coli* has three cytochrome oxidase systems. One of these has a high affinity for O₂ and functions under microaerophilic conditions, while the second has low affinity for O₂ and functions under strongly aerobic conditions (Calhoun et al. 1994). The third system (AppBC) is poorly characterized but may function as an oxygen scavenger (Dassa et al. 1991). Finally, strain EbN1 has two complete multicomponent (six or seven components each) H⁺- or Na⁺-translocating NADH:ferridoxin oxidoreductases (NFO; TC 3.D.6).

The Phosphoenolpyruvate:Sugar-Transporting Phosphotransferase System (PTS)

Strain EbN1 has a complete phosphotransferase system, but only one PTS permease. Based on homology results, this system is likely to be specific for fructose. It probably is a high-affinity system that phosphorylates the sugar at the 1-position, yielding cytoplasmic fructose-1-phosphate (Barabote and Saier 2005). It is the only recognizable sugar transporter encoded within the strain EbN1 genome. We were not able to find the IIB constituent of the fructose permease, but these small proteins are easy to miss. No nitrogen regulatory PTS phosphoryl transfer chain is present in strain EbN1 (Rabus et al. 1999).

Transmembrane Electron Flow Systems

Transmembrane oxidoreductases that transfer electrons from one side of the cytoplasmic membrane to the other without coupling proton movement to the process alter the energetics of the cell (Kimball and Saier 2002). Those which transfer electrons from in to out diminish the electrochemical potential, while those that transfer electrons from out to in increase the electrochemical potential. Strain EbN1 has several such systems. These include both a disulfide bond oxidoreductase B (electrons transferred from out to in) and a disulfide bond oxidoreductase D (electrons transferred from in to out) (see Table 2). These electron carriers are believed to function primarily in the reversible formation of disulfide bonds in extracytoplasmic (e.g., periplasmic) proteins (Nakamoto and Bardwell 2004).

Strain EbN1 has multiple prokaryotic molybdopterin-containing oxidoreductases of the PMO family (TC 5.A.3) (Rothery et al. 2008). These include (1) a nitrate reductase (Nar), (2) two potential tetrathionate reductases (Ttr), (3) a dimethylsulfoxide (DMSO) reductase (Dms), (4) a formate dehydrogenase (Fdn), (5) two polysulfide reductases (Psr), and (6) two homologues of the anaerobic ethylbenzene dehydrogenase, Ebd, one of which is already in TCDB. The

three subunits of these two enzyme complexes exhibit 74% (A), 80% (B), and 57% (C) identity, respectively, suggesting similar function. Ethylbenzene dehydrogenase is a soluble periplasmic enzyme assumed to transfer electrons, derived from anaerobic dehydrogenation of ethylbenzene, to a periplasmic c-type cytochrome (Kloer et al. 2006). Thus, it does not catalyze transmembrane electron flow as is true for several members of this family.

Auxiliary Proteins

Strain EbN1 has several auxiliary proteins that play essential roles for the normal functioning of various transporters. It has at least nine “membrane fusion proteins” that connect primary cytoplasmic active efflux pumps to outer membrane pore-forming factors (TC 1.B.17), allowing export of substrates from the cytoplasm to the extracellular medium in a single energy-coupled step (Hu and Saier 2006; Paulsen et al. 1997). Strain EbN1 also has a single system of the MPA1 family for facilitating export of polysaccharides.

As noted above, *A. aromaticum* has a single system for the transport and phosphorylation of fructose via the PTS (TC 4.A.2). To be functional, it must also possess the energy-coupling phosphoryl transfer proteins of the PTS, enzyme I and HPr. These, as well as a mannose-type IIA protein (see Table 2), were found. The three enzyme I homologues proved to resemble (1) PEP synthetases, (2) a phenylphosphate synthetase (Schleissner et al. 1994), and (3) enzyme I of the PTS. Thus, strain EbN1 has just one enzyme I. Other auxiliary proteins identified are listed in Table 2.

Incompletely Characterized Systems

Proteins in the 9 category of the TC system are either known transporters of poorly defined mechanism (9.A) or putative transporters where proof of transport function is incomplete (9.B). Only two types of transporters in the 9.A category were identified, and these are specific for divalent metal cations. PbrT homologues transport lead (Pb²⁺), while MgtE homologues transport magnesium (Mg²⁺). In neither of these cases is the mechanism of transport well defined.

We identified four FeoB (Fe²⁺ transporting) homologues (9.A.8) in strain EbN1, but all four of these give low scores with the homologues in TCDB. Moreover, characterized FeoB proteins have 8–12 C-terminal TMSs as well as an N-terminal ATP/GTP hydrolyzing domain, but the four putative FeoB homologues encoded within the genome of strain EbN1 have 0 or 1 putative TMS. Consequently they are probably G-proteins unrelated in function to Fe²⁺ transporters.

In TC category 9.B, several homologues encoded within the genome of strain EbN1 are likely to be transporters. Strain EbN1 has two putative bacterial murine precursor exporters (9.B.3), proteins that are found in virtually all bacteria. They also have two putative lipid translocases (9.B.11). All four of these homologues give good scores with TCDB entries. Two putative heme exporters (9.B.14) were identified, one with a very good score and one with a poor score. This second system may be specific for something other than heme.

Strain EbN1 encodes 21 fatty acyl CoA synthetases (FAT family; TC 9.B.17), some of which may catalyze fatty acid uptake (Zou et al. 2002). Several other putative transporters are listed in Table 2, but the transport functions of these TC entries are even less well defined than those discussed above.

Topologies of Putative Transport Proteins

Figure 2 presents the predicted topologies of the putative transport proteins encoded within the genome of strain EbN1. Many of these, including energy-coupling ATPases and phosphoryl transfer proteins, have no TMSs, while others, periplasmic solute binding receptors, targeted for secretion via the general secretory (Sec) pathway or the twin arginine translocating (TAT) system, have just one TMS. There are significantly more 6-TMS proteins than 5- or 7-TMS proteins, and more 12-TMS proteins than 11- or 13-TMS proteins; 10-TMS proteins are also more numerous than 9- or 11-TMS proteins. Although proteins with even numbers of TMSs predominate over those with odd numbers of TMSs in all organisms, the pattern shown here is quantitatively different from those observed for certain other proteobacteria such as *Bdellovibrio bacteriovorans* and other prokaryotic microorganisms (Barabote

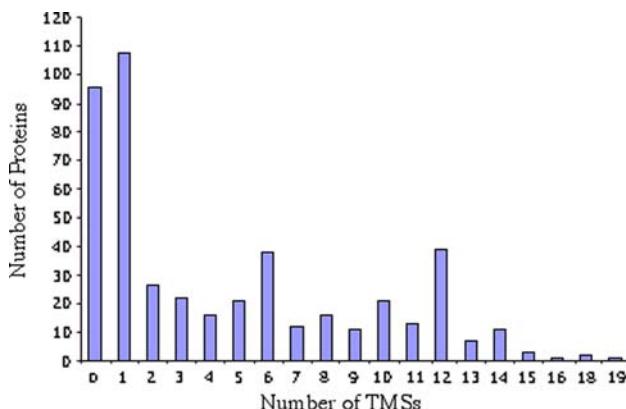


Fig. 2 Distribution of predicted topological types of proteins among the identified transporters in *A. aromaticum* strain EbN1. The numbers of proteins exhibiting a specific topological type (i.e., of a putative number of TMSs) (on the x-axis) is plotted versus the putative numbers of TMSs per polypeptide (on the y-axis)

et al. 2007; Heider and Fuchs 1997; Paulsen et al. 2000). The patterns observed must reflect the types of transport proteins that predominate. The distribution of topological types with more even-numbered TMS proteins than odd-numbered TMS proteins is not entirely surprising, as it reflects the evolutionary process (Saier 2003).

Subdivision of Strain EbN1 Transporters According to Probable Substrate Specificity

Table 3 summarizes the results presented in Table 2, subdividing the systems identified in terms of both transporter type and probable substrate specificity. Of the α -type channels, six are nonselective, six are specific for cations, and two transport anions. Of the outer membrane porins, 11 are nonspecific, although 2 prefer anionic substrates and 5 prefer cationic substrates. There are many porins that exhibit specificity for a type of substrate. One possibly recognizes sugars, while another recognizes monocarboxylates. While one takes up vitamin B₁₂ across the outer membrane, seven take up iron siderophore complexes, again in agreement with an organism that relies on electron transfer as a primary means of energy production. Four systems allow facilitation of small hydrophobic substrates such as drugs across the outer membrane, while seven function in macromolecular export (two for carbohydrates, four for proteins, and one for lipids).

Examining the primary and secondary carriers, we see some interesting parallels and differences. There are more than twofold more cation-specific carriers of both types than anion-specific carriers. In addition to the single PTS permease, there is only one potential secondary sugar transporter and no primary active transporter for sugars. However, the presence of eight monocarboxylate transporters and eight di- and tricarboxylate transporters clearly implies the importance of these exogenous metabolites as sources of energy. Amino acids and their derivatives are of moderate importance to strain EbN1, with nine primary active transporters and five secondary carriers exhibiting these specificities. Cytoplasmic membrane transporters for vitamins and cofactors are also present as expected. Macromolecular transporters, mostly exporters, are prevalent, with at least 8 for proteins, as many as 10 for carbohydrates, 6 for lipids and related compounds, and at least 2 for nucleic acids.

Transporters Specific for Hydrophobic and Aromatic Compounds

Including all types of transporters classified in TCDB, a total of 72 systems are predicted to act on hydrophobic and aromatic compounds. This corresponds to 28% of all the transporters in strain EbN1 (Table 4). This value

Table 3 Distribution of transporters based on substrate specificity

Substrate	No. of families of indicated type using substrate							Total no. of systems
	Channels	Porins	Primary carriers	Secondary carriers	Group translocator	Transmembrane electron flow carriers	(Putative) Poorly characterized	
I. Inorganic molecules								
A. Nonselective	6	10						16
B. Cations	2		21	17			6	46
C. Anions	1		6	6				13
D. Electrons						16		16
II. Carbon sources								
A. Sugars & polyols		1			1			2
B. Monocarboxylates				2			21	23
C. Di- & tricarboxylates				8				8
D. Organoanions (noncarboxylic)		2						2
E. Aromatic compounds				10				10
III. Amino acids & their derivatives								
A. Amino acids & conjugates		3	4					7
B. Amines, amides, polyamines, & organoacations	2			1				3
C. Peptides		4	1			2		7
IV. Vitamins, cofactors, & cofactor precursors								
A. Vitamins & vitamin or cofactor precursors		1						1
B. Enzyme & redox cofactors		1				2		3
C. Siderophores; siderophore-Fe complexes		8						8
V. Drugs, dyes, sterols, & toxics								
A. Multiple drugs	5		16					21
B. Specific drugs		9						9
C. Pigments								
D. Other hydrophobic substances		4	9					13
VI. Macromolecules								
A. Carbohydrates	2	4	1			4		11
B. Proteins	5	9	3					17
C. Nucleic acids		2						2
D. Lipids	1	2	2			1		6
VII. Unknown								
Total	11	32	68	84	1	16	46	258

corresponds to the largest percentage yet recorded for a living organism, to the best of our knowledge (see Fig. 3). Among these, 10% to 13% are predicted to be specific for aromatic compounds. Again, this is a higher percentage than observed for most bacteria. These values are consistent with expectation for an organism that makes its living eating hydrophobic and aromatic substances.

Discussion

A. aromatoleum strain EbN1 is best known for its ability to degrade hydrophobic and aromatic substances. We examined the genome of strain EbN1 for transporters likely to be specific for these types of compounds (Table 4) and found 9 uptake transporters (3%) for small aromatic compounds,

Table 4 Transporters specific for hydrophobic and aromatic compounds

Compound	TC no.	TC homologue	Number	Total
Hydrophobic/amphipathic compounds				
	2.A.1.2	DHA1	4	32
	2.A.1.3	DHA2	1	
	2.A.6.2	HAE1	9	
	2.A.6.7	HAE3	7	
	2.A.6.8	ORF4	1	
	2.A.7.1	SMR	1	
	2.A.66.1.1	MATE	1	
	3.A.1.105	DrugE1	2	
	3.A.1.120	DrugRA1	4	
	3.A.1.121	DrugRA2	1	
	3.A.1.122	MacB	2	
	3.A.1.201	MDR	1	
Hydrophobic amino acids	3.A.1.4.1	ABC HAAT	3	3
Fatty acids/lipids	1.B.9.1.1	FndL	1	27
	2.A.1.42.1	LpIT	2	
	3.A.1.106	LipidE	1	
	3.A.1.114	DevE	1	
	9.B.11.1.1	Tgd1	1	
	9.B.17.1.4	FadD	15	
	9.B.17.1.6	CaiC	6	
Aromatic compounds	2.A.1.15	AAHS	3	10
	2.A.1.27	PPP	1	
	2.A.1.30	ADT	1	
	2.A.1.36	YnfM	1	
	2.A.21.7.1	Ppa	1	
	2.A.71.2.1	FBT	1	
	5.A.3.9	PMO	2	
Total no. of transporters for hydrophobic & aromatic compounds				72

30 efflux transporters (11%) for hydrophobic/amphiphilic drug-like substances, and 5 (~2%) specific for the uptake of hydrophobic amino acid-like compounds. It should be recalled that these systems might show broad specificities. In particular, the putative drug exporters are likely to be able to act on a variety of aromatic and hydrophobic compounds encountered by strain EbN1 in the natural environment, particularly at contaminated sites.

The uptake systems for aromatic compounds could be of broad specificity, transporting a variety of aromatic toxins. The five ABC hydrophobic amino acid uptake systems probably transport aromatic amino acids and other hydrophobic/aromatic substances. The high percentage of transporters for aromatic and hydrophobic substances correlates with the very low percentage of transporters for sugars (at most two systems, which may not be complete) and hydrophilic or semipolar amino acids (five systems: three uptake and two efflux systems).

Although ABC uptake systems are sparse in strain EbN1 compared to many other bacteria, ABC efflux systems are

more numerous. In addition to protecting cells against toxic substances, ABC efflux pumps export periplasmic, surface, and extracellular macromolecules: carbohydrates, lipids, and proteins. Surprisingly, four of the ABC efflux systems in strain EbN1 more closely resemble characterized eukaryotic systems than known prokaryotic systems. All of these are half-sized pumps which presumably function as dimers. One of the two heavy metal efflux pumps (3.A.1.210) is 50% larger than its nearest eukaryotic homologue. The system that resembles a mitochondrial peptide exporter (3.A.1.212) may have shared a common ancestry with it, both having diverged from each other during vertical descent.

An ABC transporter with an extra C-terminal 300 hydrophilic residues (EbA6882) proved to have an unusual domain structure as revealed by screening the conserved domain database (CDD). The extra 300 residues contained two adjacent domains, a histidine kinase-like ATPase (HATPase-c) domain and a signal receiver (REC) domain, common to all response regulators. The latter are

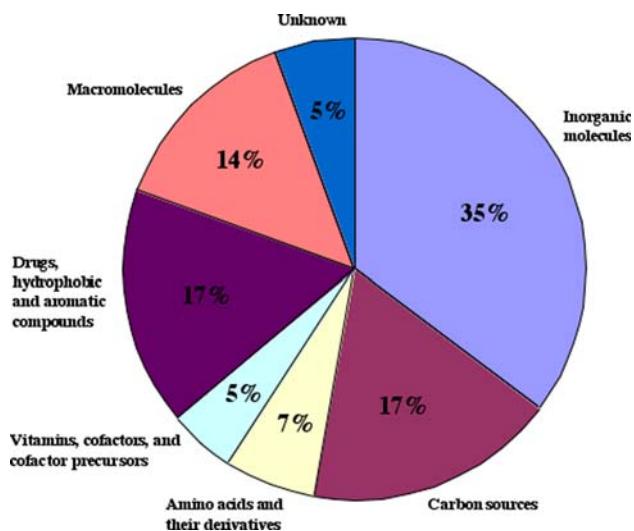


Fig. 3 Estimates of the percentages of recognized transport systems found in *A. aromaticum* sp. EbN1 according to substrate type. Inorganic molecules are primarily ionic species and small, hydrophilic, neutral molecules such as H_2O , NH_3 , and CO_2 . Macromolecules include proteins, complex cell surface carbohydrates, and lipids. Other categories are self-explanatory

phosphorylated by histidine kinases. Three possibilities therefore can be considered concerning their mode of regulation: (1) phosphorylation of this domain may regulate the ABC transport activity, (2) the transporter could provide a signal for control of regulatory phosphoryl transfer via the C-terminal domains, and (3) the transport and regulatory components may function independently. A BLAST search revealed that this fusion arrangement, with an ABC transporter fused to HATPase and REC domains, is only found in a few β -proteobacteria closely related to strain EbN1 (species of *Ralstonia* and *Burkholderia*). These fusion proteins have not previously been investigated. They may provide a novel regulatory mechanism.

Strain EbN1 has a huge variety of macromolecular transport systems. It has a full complement of proteins for assembly of (1) the general secretory (Sec) pathway; (2) the Oxa system, which can insert proteins into the cytoplasmic membrane dependently and independently of the Sec system; (3) the twin arginine targeting and translocating (Tat) system; (4) the outer membrane protein insertion porin (OmpIP) complex; (5) three type I ABC protein secretion systems; (6) two type IIb (MTB)-type outer membrane protein secretion systems, one of the Pul type for general protein secretion and one of the Pil type for pilus subunit secretion; (7) a type IV conjugation-type (Conj) secretion system for export of DNA-protein complexes involved in conjugation; and (8) a septal DNA translocator (S-DNA-T). It also has numerous transporters for lipids, fatty acids, lipoproteins, and complex carbohydrates.

The results described here provide a comprehensive overview of the transport capability of *A. aromaticum* strain EbN1. They should serve as guides for future research concerning the ecophysiological role of this organism in its natural habitat and assessment of bioremediation prospects. Strain EbN1 can be regarded as an aquatic degradation specialist feeding on low molecular weight compounds ranging from plant exudates (e.g., dicarboxylic acids) to naturally as well as anthropogenically released toxic aromatic compounds (e.g., toluene and phenol). The present study reveals that the transporter complement of strain EbN1 matches its degradative capacities, whereby a well-integrated system is established for uptake and complete oxidation (to CO_2) of organic growth substrates. Notably, strain EbN1 contains paralogous transport systems in several cases, agreeing with the high genome plasticity of strain EbN1. One may speculate that these systems possess different ranges of substrate specificity and/or affinity, allowing strain EbN1 to adapt its transporter equipment to changing substrate concentrations in the natural environment. Considering the high degree of substrate-dependent regulation of catabolic pathways in this organism (Kühner et al. 2005; Wöhlbrand et al. 2007, 2008), similar regulatory control mechanisms can be envisioned for some of the transporters defined in the present *in silico* analysis.

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